

Analysis of Cell Culture Amino Acids in a Supplement Using the Alliance™ iS HPLC System and AccQ Tag™ Ultra Derivatization

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Abstract

Amino acid analysis (AAA) is fundamental to nutritional research and dietary supplement quality assessment, yet it remains analytically challenging. Many amino acids lack strong chromophores, requiring derivatization to enable sensitive detection. AccQ•Tag Ultra Derivatization Chemistry is a widely adopted and highly robust solution to these challenges. The pre-column derivatization method employs 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate (AQC), a reagent that rapidly and quantitatively labels both primary and secondary amines to form stable derivatives. When paired with reversed-phase chromatography, this chemistry delivers high sensitivity, excellent stability, and reproducible retention times, making it particularly well-suited for complex dietary supplement matrices.

In this study, the performance of the Alliance iS HPLC System in combination with AccQ•Tag Ultra Amino Acid Analysis Consumables will be assessed for quantitation of dietary supplements. The system supports optimized AccQ•Tag Chemistry workflows with 2.5 µm columns, providing precise quaternary gradient delivery, controlled column temperature, and sensitive UV detection. This approach will show the benefits of combining Alliance iS

HPLC System and AccQ•Tag Column chemistry with advanced system intelligence, making it a strong choice for laboratories performing routine amino acid analysis in dietary supplement testing.

Benefits

- Alliance iS HPLC System in combination with AccQ•Tag Ultra 2.5 µm Columns, Reagents, and Consumables provides a reliable, robust method for HPLC analysis of both hydrolysates and cell culture amino acids, designed for regulated laboratories.
- The combination of system and consumables with Empower™ 3 Chromatography Data System (CDS) provides a turnkey approach for amino acid analysis of dietary supplements, allowing regulated laboratories to benefit from a modern, compliance-ready system, designed to reduce common setup mistakes.

Introduction

AAA is a foundational technique in analytical chemistry, supporting essential workflows in biopharmaceutical development, nutritional science, and cell culture optimization. In nutritional research, precise amino acid profiling informs dietary formulation, quality control, and food composition studies, making accurate and reproducible analytical methods critical for both clinical and industrial applications. Among available approaches, the AccQ•Tag Ultra Chemistry—combining the AQC derivatization reagent with dedicated columns and eluents—has become a well-established methodology due to its exceptional sensitivity, robustness, stability, and reproducibility. The method uses pre-column derivatization with 6-aminoquinolyl-N-hydroxysuccinimidyl carbamate (AQC), which reacts quantitatively with both primary and secondary amines to produce highly stable fluorescent derivatives.

The reversed-phase separation of AQC-labeled amino acids with AccQ•Tag Ultra Columns and Reagents has demonstrated excellent reproducibility on the ACQUITY Arc™ System and Arc HPLC System.¹ Achieving consistent performance requires an LC platform capable of delivering precise quaternary gradients, maintaining tight temperature control in the column compartment, and providing highly sensitive UV detection. In the following study, the method was transferred to the Alliance iS HPLC System with TUV detection for the analysis of amino acids in dietary supplements, demonstrating its suitability for regulated laboratories that require quantitation of amino acids.

Experimental

Sample Description

All calibration standards were prepared from Waters Amino Acid Cell Culture Standard Kit (p/n: [186009300 < https://www.waters.com/nextgen/global/shop/standards--reagents/186009300-amino-acid-cell-culture-standard-kit.html>](https://www.waters.com/nextgen/global/shop/standards--reagents/186009300-amino-acid-cell-culture-standard-kit.html)) using norvaline (p/n: [186009301 < https://www.waters.com/nextgen/global/shop/standards--reagents/186009301-amino-acid-internal-standard-norvaline.html>](https://www.waters.com/nextgen/global/shop/standards--reagents/186009301-amino-acid-internal-standard-norvaline.html)) as the internal standard and 0.1 N hydrochloric acid (HCl) in water as the diluent.³ The internal standard stock was prepared at 2500 µM in 0.1 N HCl. The final concentration of the calibrants varied to match the content range of each supplement, and a concentration of 250 µM was used for norvaline (internal standard).

Amino Acid Supplement Capsule Sample Preparation

The amino acid supplement capsule sample (past expiry) was collected and weighed. Three capsules represent one serving; therefore, providing an adequate sample size for uniformity, nine capsules and three servings were prepared. The final weight of the nine capsules was 7255.76 mg. The sample was then diluted in 200 mL of 0.1 N HCl in water. The sample was then sonicated for 1.5 hours followed by stirring for 1 additional hour. The sample was filtered using a 0.45 µm HA filter.

An additional dilution of 1:50 was performed on the sample extraction, making the final dilution of the sample 1:10,000. The sample was then derivatized using Waters AccQ•Tag Ultra Derivatization Protocol.⁴

LC Conditions

LC system:	Alliance iS HPLC System
Detection:	TUV Detector
Wavelength:	260 nm
Sampling rate:	10 Hz
Vials:	LCGC Certified Clear Glass 12 x 32 mm Screw

	Neck Vial, total recovery with cap and PTFE/Silicone septum (not pre-slit) (p/n: 186000384C)
Column(s):	AccQ•Tag Ultra C18, 2.5 µm 4.6 x 150 mm Column (p/n: 186010407)
Column temperature for hydrolysate and cell culture standard:	43 °C
Sample temperature:	20 °C
Injection volume:	2 µL
Flow rate:	1.5 mL/min
Mobile phase A:	AccQ•Tag Ultra Eluent A (p/n: 186003838)
Mobile phase B:	90:10 (v/v) Water:AccQ•Tag Ultra Eluent B
Mobile phase C:	Milli-Q Water
Mobile phase D:	AccQ•Tag Ultra Eluent B (p/n: 186003839)
Sample manager wash:	95:5 (v/v) Water:Acetonitrile
Sample manager purge:	95:5 (v/v) Water:Acetonitrile

Gradient Table

Time (min)	Flow (mL/min)	%A	%B	%C	%D	Curve
Initial	1.500	10.0	0.0	90.0	0.0	Initial
0.36	1.500	10.0	0.0	90.0	0.0	11
19.67	1.500	9.0	80.0	11.0	0.0	7
25.07	1.500	8.0	16.0	60.0	16.0	7
25.74	1.500	8.0	16.0	58.0	18.0	6
27.05	1.500	7.8	0.0	70.9	21.3	6
28.20	1.500	4.0	0.0	36.3	59.7	6
30.08	1.500	4.0	0.0	36.3	59.7	6
30.38	1.500	10.0	0.0	90.0	0.0	6
35.48	1.500	10.0	0.0	90.0	0.0	6

Data Management

Chromatography data system:

Empower 3.9.0 CDS

Results and Discussion

Amino Acid Profile of a Dietary Capsule Supplement Sample

To assess the robustness of the method, the method that was previously verified on the ACQUITY Arc HPLC System¹ was transferred to the Alliance iS HPLC TUV System. For this testing, the cell culture standard was evaluated on the Alliance iS HPLC System for repeatability, linearity, and sensitivity. Specific parameters tested included limits of detection and quantitation (LOD/LOQ), calibration linearity, and intraassay precision, producing comparable results as previously observed.¹ These findings in Table 1 indicate that the Alliance iS HPLC System provides consistent performance to support robust amino acid analyses, meeting method criteria to support quantitation. The chromatographic separation for amino acid hydrolysate and cell culture standards are shown in the stacked chromatogram overlay in Figure 1.

Parameter	Results at 20 pmol
Retention Time %RSD	0.00–0.02
Area %RSD	0.50–0.78
USP Tailing	<1.2
USP Resolution	<ul style="list-style-type: none"> ➤ 2.1, critical pair Arg/Gly ➤ All other peaks >2.3
Linearity (5–500 μM)	$R^2 \geq 0.998$ all peaks, <10% residuals for low calibrants, all other points <7%

Table 1. Cell culture standard method performance on Alliance iS HPLC System.

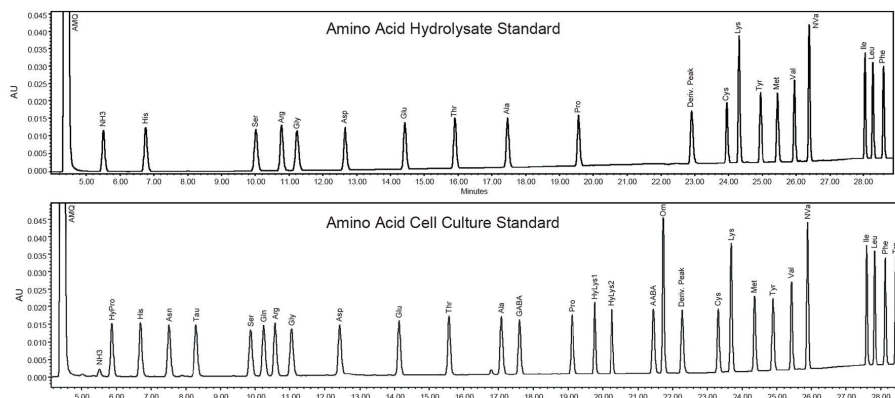


Figure 1. Amino acid hydrolysate standard chromatogram and amino acid cell culture standard chromatogram analyzed on Alliance iS HPLC TUV System.

Following evaluation of the method performance, commercial amino acid capsule supplements were analyzed to quantitatively determine their amino acid content relative to manufacturer label claims. Given the complex

sample matrix, the supplements were extracted prior to derivatization. Due to the complexity of the sample components, some optimization of sample preparation was conducted. The amino acid derivatives in the matrix were effectively resolved using reversed-phase liquid chromatography under optimized gradient conditions, providing consistent retention and peak shape across the analyte panel.¹ No interference peaks were observed. The amino acid capsule supplement sample and amino acid cell culture standard is shown in Figure 2.

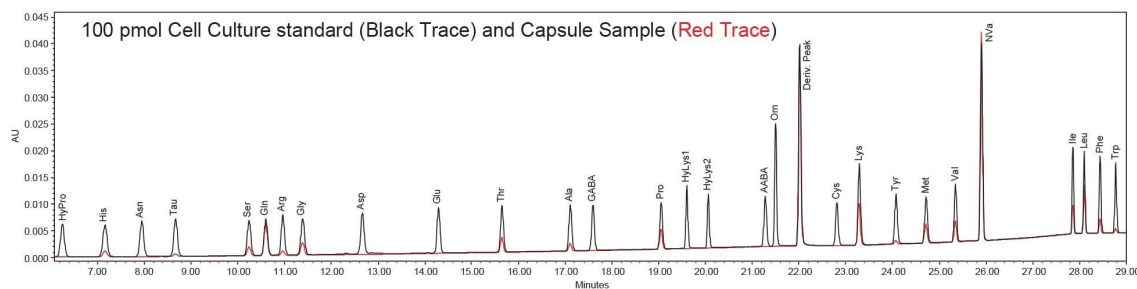


Figure 2. Zoomed in chromatogram overlay of the amino acid cell culture standard and amino acid capsule sample on Alliance iS TUV HPLC System.

Quantitative results were compared with declared label values to assess accuracy and compliance. This type of assessment is essential for verifying product integrity, supporting quality assurance processes, and aligning with regulatory expectations for dietary supplement labeling. The recovery data demonstrated that the analytical workflow—AQC derivatization combined with RPLC separation on the Alliance iS HPLC System—delivered reliable performance across all amino acids assessed. Recoveries ranged from 75% to 113%, with the majority falling within the preferred 88–99% range, indicating strong agreement with expected concentrations. These results are consistent with typical performance for amino acid analysis in complex supplement matrices, where extraction efficiency and matrix interactions often influence final recoveries.

Compound	Amount in Sample Label Claim Per serving (mg)	Amount in Quantitated Per serving (mg)	% Recovery
Histidine	231	203	87.9
Taurine	69	61	88.2
Serine	204	201	98.6
Glutamine	1014	947	93.4
Arginine	204	153	75.1
Glycine	204	194	92.3
Threonine	312	292	93.6
Alanine	123	115	93.5
Proline	405	379	93.7
Lysine	675	568	84.1
Tyrosine	108	118	109.8
Methionine	474	470	99.1
Valine	339	320	94.4
Isoleucine	351	350	99.7
Leucine	621	615	99.1
Phenylalanine	258	285	110.5
Tryptophan	99	112	113.2

Table 2. Amino acid content in amino acid capsule supplement sample for amount prepared (nine capsules) including quantitated percent (%) recovery results.

A few amino acids exhibited recoveries near the upper or lower bounds of the observed range. These deviations may arise from incomplete extraction, matrix suppression or enhancement, or inherent variability in derivatization efficiency among certain amino acids. Although these results remain within acceptable limits for dietary supplement analysis, further optimization of the extraction procedure—such as evaluating alternative solvent systems, adjusting extraction time, or incorporating additional mechanical agitation—may help improve recoveries.

Overall, the tight clustering of recoveries for most amino acids reflects the robustness of both the derivatization chemistry and the chromatographic method. AQC reagent chemistry is well characterized for its stability and efficient derivatization of diverse amino acids, enabling reproducible fluorescence based or UV based detection. The Alliance iS HPLC System further enhanced method reproducibility through intelligent instrument monitoring, stable flow control, and environmental consistency, reducing variability during routine operation.

Conclusion

Accurate quantification of amino acids in dietary supplements is essential for verifying label claims, ensuring product integrity, and meeting regulatory expectations for nutritional product labeling and quality control. Because amino acids serve as key functional ingredients, analytical methods must exhibit high precision, accuracy, and robustness to satisfy product quality.

In this study, the combination of AQC precolumn derivatization and reversed-phase HPLC separation on the Alliance iS HPLC System produced strong analytical performance across a diverse amino acid panel. Percent recoveries ranged from 75% to 113.2% relative to manufacturer label claims, with most amino acids falling within the preferred 88%–99% range. These results highlight the effectiveness of the AccQ•Tag Ultra Derivatization Chemistry as well as the chromatographic stability, reproducibility, and overall reliability of the Alliance iS HPLC System.

Overall, the precision and recovery results validate the robustness of the analytical workflow and confirm the suitability of the Alliance iS HPLC System for routine quality control applications in regulated laboratory environments. The strong alignment between measured concentrations and declared label values supports product integrity and reinforces confidence in the method's quantitative accuracy. By integrating advanced intelligent instrumentation with rigorously validated chromatographic methodologies, the Alliance iS HPLC System enables laboratories performing amino acid analysis to maintain analytical robustness, achieve reproducible data quality, and uphold regulatory compliance without sacrificing operational efficiency or throughput.

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