

## Benchmarking Resolution and Recovery of BioResolve™ 1 mm ID C<sub>18</sub> RP Columns with MaxPeak™ Premier Technology

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### Abstract

The BioResolve Peptide C<sub>18</sub> RP 1.0 mm ID Column with MaxPeak Premier Technology was benchmarked against 3 commercially available microflow columns of similar dimensions. Using a MassPREP™ Enolase Digest with Phosphopeptides Mix, the BioResolve Peptide C<sub>18</sub> RP Column demonstrates superior peak capacity, improved peak shape, and significantly higher recovery of acidic and phosphorylated peptides from the first injection. As a result, the BioResolve Peptide C<sub>18</sub> RP Column delivers reproducible separations and exceptional mass spectral data quality, improving confidence in peptide identification. These results highlight the advantages of MaxPeak Premier Technology in microflow proteomics workflows and underscore the BioResolve Peptide C<sub>18</sub> RP Column's ability to deliver sensitive, reproducible separations without extensive column conditioning.

### Benefits

- Exceptional recovery and peak shape of acidic and phosphorylated peptides from the first injection, eliminating the need for extensive column conditioning.

- Higher peak capacity compared to other commercially available 1.0 mm ID columns, enhancing separation performance.
  - Enhanced mass spectrometry (MS) sensitivity and cleaner spectra, improving confidence in peptide identification.
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## Introduction

Microflow chromatography offers several advantages over traditional analytical scale chromatography in proteomics applications. Microbore columns require less sample load and enable the use of low flow rates (10–100  $\mu\text{L}/\text{min}$ ). These low flow rates decrease solvent consumption and optimize the interface with MS, increasing MS sensitivity.<sup>1,2</sup> However, recovery of acidic peptides in microflow proteomics workflows can be challenging due to non-specific adsorption to stainless steel column hardware.

Waters 1.0 mm ID MaxPeak Premier Columns use MaxPeak High Performance Surfaces (HPS) Technology to mitigate unwanted interactions between acidic analytes and column hardware.<sup>3,4</sup> BioResolve Peptide 1.0 mm ID Columns with MaxPeak Premier Technology demonstrate excellent recovery and peak shape for acidic peptides from the first injection, eliminating the need for extensive column conditioning.<sup>4</sup> Here, BioResolve Peptide C<sub>18</sub> RP 1.0 mm ID Columns are benchmarked against 3 commercially available 1.0 and 1.5 mm ID columns. The BioResolve Peptide Column outperforms the alternative columns in peak shape and recovery of highly acidic peptides, resulting in a cleaner baseline and improved signal-to-noise from the first injection.

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## Experimental

### Sample Description

Waters MassPREP Enolase Digest with Phosphopeptides Mix (p/n: [186003286 < https://www.waters.com/nextgen/global/shop/standards--reagents/186003286-massprep-enolase-digest-with-phosphopeptides-mix.html>](https://www.waters.com/nextgen/global/shop/standards--reagents/186003286-massprep-enolase-digest-with-phosphopeptides-mix.html) ) was reconstituted in 100  $\mu\text{L}$  of 0.1% formic acid in water.

### Peptide Sequences

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Peptide	Sequence
T35	WLTGPQLADLYHSLMK
T37	YPIVSIEDPFAEDDWEAWSHFFK
T51	IEEELGDNAVFAGENFHHGDK
T51-52	IEEELGDNAVFAGENFHHGDKL
T18p	NVPLpYK
T19p	HLADpSK
T43p	VNQIGpTLSESIK
T43pp	VNQIGpTLSEpSIK

## Calculations

4 $\sigma$  peak capacities were determined based on the following equation:

$$P_{C,4\sigma} = 1 + \left[ \left( \frac{2.35}{4} \right) \left( \frac{t_{gradient}}{W_{h,avg}} \right) \right]$$

## LC Conditions

LC system and setup:

ACQUITY™ Premier UPLC™ Standard tubing prior to column, flow diverted straight from the column to the MS source with 40  $\mu$ m ID x 30 in tubing (p/n: 700011516).

Columns:	BioResolve Peptide C <sub>18</sub> RP Column, MaxPeak Premier Technology, 1.7 μm, 130 Å, BEH, 1.0 x 100 mm (p/n: 186011505) "Column K": core-shell C <sub>18</sub> 1.7 μm, 100 Å, 1.0 x 100 mm "Column Y": C <sub>18</sub> 1.9 μm, 120 Å, 1.0 x 100 mm "Column H": core-shell C <sub>18</sub> 2.7 μm, 120 Å, 1.5 x 100 mm
Column temperature:	60 °C
Sample temperature:	6 °C
Injection volume:	1 μL
Mobile phase A:	0.1% Formic Acid in H <sub>2</sub> O
Mobile phase B:	0.1% Formic Acid in ACN
Sample vials:	QuanRecovery™ MaxPeak 12 x 32 mm Propylene 300 μL Screw Cap Vials (p/n:186009186)

## Xevo™ G2 Detector Settings

ESI probe:	Low-flow probe (p/n: 186007529)
Mass range:	50–2000 <i>m/z</i>
Mode:	ESI+
Sample rate:	1 Hz

Cone voltage:	30 V
Source temperature:	120 °C
Desolvation temperature:	250 °C
Capillary voltage:	2.50 kV
Cone gas:	50 L/h
Desolvation gas:	350 L/h
Lockmass:	LeuEnk (556.27658 <i>m/z</i> )
Informatics:	waters_connect™ Software

### Gradient Table (Waters Column, Column K, Column Y)

Time (min)	Flow (μL/min)	%A	%B	Curve
Initial	100	99	1	Initial
18.00	100	60	40	6
18.50	100	15	85	6
19.40	100	15	85	6
20.30	100	99	1	6
25.30	100	99	1	6

## Gradient Table (Column H, conditions adjusted to the column ID of 1.5 mm)

Time (min)	Flow (μL/min)	%A	%B	Curve
Initial	142	99	1	Initial
21.22	142	60	40	6
21.81	142	15	85	6
22.88	142	15	85	6
23.94	142	99	1	6
29.83	142	99	1	6

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## Results and Discussion

Figure 1 shows the total ion chromatograms (TICs) from the first injection of the MassPREP Enolase Digest with Phosphopeptides Mix on a BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors. Column K and Column Y have the same dimensions as the BioResolve Peptide C<sub>18</sub> RP Column (1.0 x 100 mm), while Column H has a larger inner diameter (1.5 x 100 mm). Separation performance for non-acidic peptides was quantified for each column by calculating the average peak capacity across 4 peptides. Of the three 1.0 mm ID columns tested, the BioResolve Peptide C<sub>18</sub> RP Column exhibits the best separation performance for non-acidic peptides; its peak capacity is over 10% higher than Column K and Column Y. A slightly higher peak capacity is observed on Column H, as expected due to the column's larger inner diameter which reduces the deleterious impact of post-column band dispersion. However, the BioResolve Peptide C<sub>18</sub> RP Column far outperforms all three alternative columns in the separation of acidic peptides, leading to improved reproducibility and signal-to-noise, as discussed in more detail below.

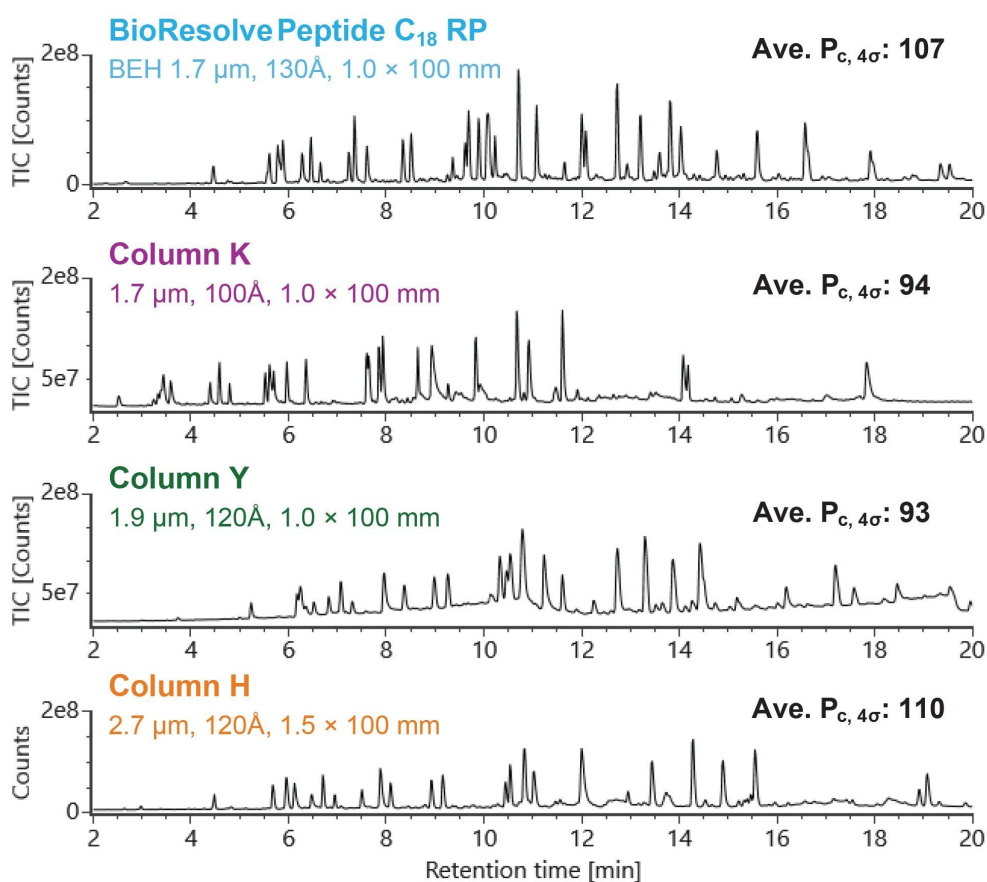


Figure 1. TICs of the MassPREP Enolase Digest with Phosphopeptides Mix on a Waters BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors. Data from the first injection on each column is shown. Average peak capacities calculated from 4 non-acidic peptides are shown on each chromatogram. Column Y exhibits a high baseline, attributed to column bleed of species with predominant mass-to-charge ratios of 149.04 m/z and 223.06 m/z.

## Peak Shape and Recovery of Acidic and Phosphorylated Peptides

Extracted ion chromatograms (XICs) of 4 phosphorylated peptides from the MassPREP Enolase Digest with Phosphopeptides Mix on a BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors are shown in Figure 2. Data from the first injection on each column is shown. The BioResolve Peptide C<sub>18</sub> RP Column uses MaxPeak Premier hardware that mitigates secondary interactions with acidic analytes. As a result, all 4

phosphopeptides are recovered on the first injection, including the doubly phosphorylated T43pp peptide. T43pp is poorly recovered on the first injection on Column K and Column Y and not recovered on Column H. The T34pp peak area is plotted against injection number for each column on the top right of Figure 2. The T43pp peak area on the BioResolve Peptide C<sub>18</sub> RP Column plateaus by injection 3 while Column K, Column Y, and Column H show an increase in peak area with each injection. The T43pp peak area on injection 5 on all 3 alternative columns is lower than that of injection one on the BioResolve Peptide C<sub>18</sub> RP Column.

On all columns, the phosphopeptides exhibit peak tailing on the first injection and improved peak shape after column conditioning. Notably, the BioResolve Peptide C<sub>18</sub> RP Column exhibits the least phosphopeptide peak tailing of the 4 columns studied. The USP tailing factor for the T19p peptide is plotted against injection number in the bar chart on the bottom right of Figure 2. Even with conditioning, the T19p tailing factor is higher on Column K and Column Y than on the unconditioned BioResolve Peptide C<sub>18</sub> RP Column. Column H exhibits the highest T19p tailing factor without conditioning and required significant column conditioning for improved peak shape.

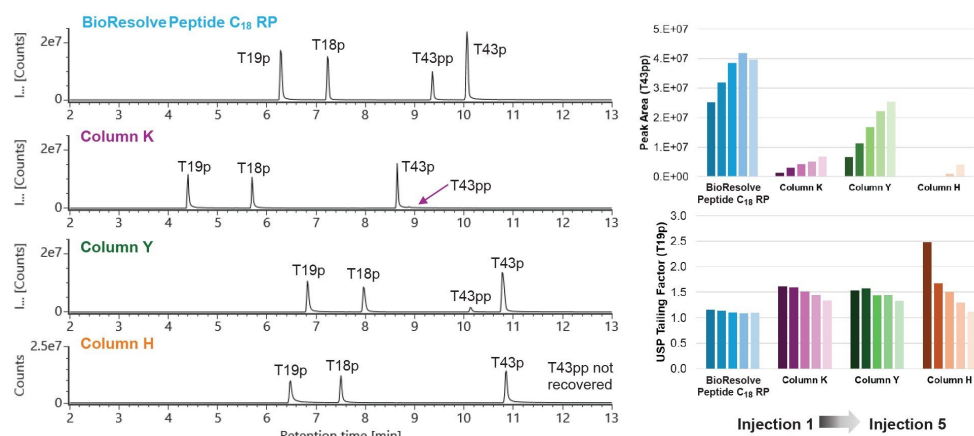


Figure 2. XICs of 4 phosphopeptides from the MassPREP Enolase Digest with Phosphopeptides Mix on a Waters BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors. Data from the first injection on each column is shown. Bar charts depicting the peak area of the T43pp peptide (top) and the USP tailing factor of the T19p peptide (bottom) are shown on the right. Amino acid sequences for each peptide are listed in the experimental section.

Figure 3 shows the XICs of 4 acidic peptides from the MassPREP Enolase Digest with Phosphopeptides Mix on a



Waters BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors. Data from injections 1, 3, and 5 are shown. All 4 acidic peptides elute as sharp peaks with minimal tailing on the first injection on the BioResolve Peptide C<sub>18</sub> RP Column. On Column Y, peak tailing is observed for all 4 acidic peptides, resulting in low peak intensities relative to the BioResolve Peptide C<sub>18</sub> RP Column. Injections 3 and 5 exhibit a slight improvement in peak tailing attributable to the column conditioning, but peak intensities are still over 40% lower on the conditioned Column Y than on the unconditioned BioResolve Peptide C<sub>18</sub> RP Column. Column K and Column H exhibit very poor separation of the 4 acidic peptides as shown in Figure 3; peaks are wide, asymmetric, and exhibit low signal intensity on the first injection, likely due to secondary interactions with the column hardware. Peak shape improves with column conditioning, but substantial tailing is still observed on injection 5. These results underscore the superior recovery and peak shape of acidic and phosphopeptides on the BioResolve Peptide C<sub>18</sub> RP Column and demonstrate the reduced need for column conditioning relative to alternative columns.

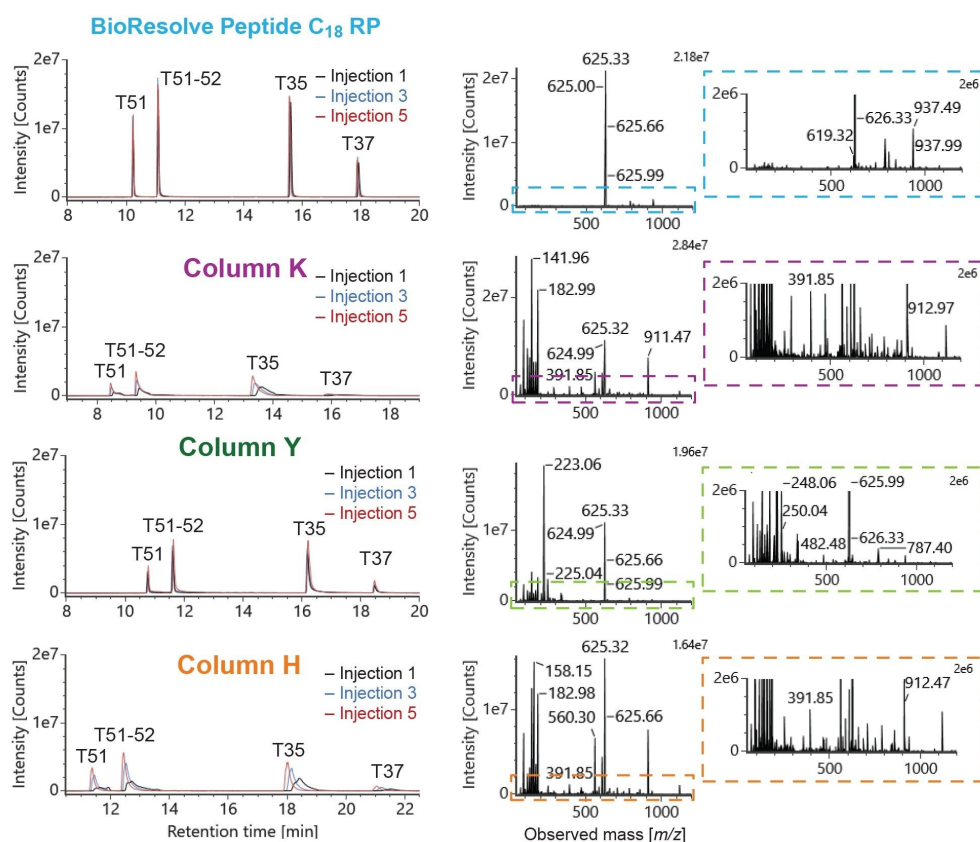


Figure 3. XICs of 4 acidic peptides from the MassPREP Enolase Digest with Phosphopeptides Mix on a Waters BioResolve Peptide C<sub>18</sub> RP Column and 3 columns from alternative vendors. Data from injections 1, 3, and 5 are shown. The mass spectra for the T35 peptide from the first injection for each column are shown on the right. Amino acid sequences for each peptide are listed in the experimental section.

## Impact of Separation Performance on MS Data Quality

The superior separation of acidic and phosphorylated peptides on the BioResolve Peptide C<sub>18</sub> RP Column improves the quality of mass spectral data, enhancing confidence in peptide identification. The mass spectra for the T35 peptide from the first injection on each column are shown on the right of Figure 3. The cleanest spectrum is obtained on the BioResolve Peptide C<sub>18</sub> RP Column, exhibiting the highest signal for the triply charged ion (625.33 *m/z*) with minimal noise from co-eluting species. On all 3 alternative columns, the signal for the triply charged T35 ion is over 40% lower than the BioResolve Peptide C<sub>18</sub> RP Column. The T35 spectra from

Columns K and H show high baseline noise across the entire mass-to-charge range analyzed. Both columns exhibit wide, asymmetric acidic peptide peaks, leading to co-elution of multiple species and poor signal-to-noise in the mass spectra. Less noise is exhibited in the T35 mass spectra from Column Y, but a high signal at 223.06  $m/z$  is observed due to column bleed. These results demonstrate the impact of chromatographic separation on MS signal-to-noise, highlighting the benefits of the improved separation of the BioResolve Peptide C<sub>18</sub> RP Column in proteomics workflows.

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## Conclusion

The BioResolve Peptide C<sub>18</sub> RP 1.0 mm ID Column with MaxPeak Premier Technology provides clear performance advantages for microflow proteomics applications. Its enhanced surface chemistry minimizes secondary interactions with acidic analytes, enabling sharper peak shapes, higher recovery, and excellent reproducibility from the first injection compared to currently available column technologies. Head-to-head comparison to three alternative microflow columns demonstrates higher peak capacity, improved recovery and peak shape of acidic and phosphorylated peptides, and cleaner MS data on the BioResolve Peptide C<sub>18</sub> RP Column. These columns offer a robust and sensitive LC-MS proteomics solution that is both economical and environmentally sustainable.

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