

Note d'application

Cracking the Challenge: Automated Extraction of PFAS in Backyard and Store-Bought Cage-Free Eggs Using LC-MS/MS and Dual-Phase GCB/WAX Cartridges

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Abstract

Many consumers enjoy eating eggs from backyard chickens and remain unaware of the environmental and dietary factors that influence egg safety. Those who consume eggs from backyard chickens often believe their eggs are healthier, yet they may be unknowingly exposing themselves to per- and polyfluoroalkyl substances (PFAS) contamination from sources like the environment, bedding, food, and drinking water. This study compares PFAS concentrations in store-bought cage-free eggs and cage-free eggs from backyard chickens, which may be exposed to a broader range of environmental factors and dietary variations due to free-range access and consuming kitchen scraps. A workflow for the analysis of whole egg (a dense, proteinaceous, and fatty matrix) is presented, utilizing automated sample preparation to reduce analyst involvement, minimize variability, and improve robustness when handling this challenging matrix. The automated sample extraction process takes less than 15 minutes per sample, and the automated solid-phase extraction (SPE) system can process up to 8 samples simultaneously in under 70 minutes. This method also uses dual-phase Waters™ Oasis™

GCB/WAX for PFAS Analysis Cartridges. The graphitized carbon black (GCB) and weak anion exchange (WAX) SPE cartridges clean up challenging samples to ensure precise and repeatable results across samples. Ultimately, this study aims to provide a clearer understanding of PFAS contamination in both commercially and locally sourced eggs, contributing insights into food safety through the creation of a consistent and efficient automated method for extracting PFAS from the challenging matrix of whole raw egg.

Benefits

- A streamlined and automated PFAS workflow for sample preparation of a complex matrix (eggs) with sensitive analysis on the Xevo™ TQ Absolute Mass Spectrometer to quantify at the sub-ng/g levels required to meet the guidance and regulations from the FDA and EU
- Use of dual-phase Oasis PFAS GCB/WAX Cartridges, combining 2 sorbents in one device, to streamline SPE, ensure cleanliness, and reduce false positives through QC-release testing for low residual PFAS
- The PFAS Solution Installation Kit reduces the risk of system and solvent contamination providing confidence in the accuracy of results

Introduction

PFAS are known as potential contaminants in eggs, which are commonly consumed by humans. Chickens are exposed to PFAS through various routes, such as their diet, bedding, environment, and drinking water, with free-ranging chickens likely exposed to more variation in those factors.^{1, 3, 6} Unlike the European Union (EU), which has set maximum levels for 4 PFAS compounds (PFOS, PFOA, PFNA, and PFHxS)², the United States currently has no federally established tolerable intake limits for PFAS in food. The European Food Safety Authority (EFSA) has also established a tolerable weekly intake (TWI) of 4.4 ng/kg body weight for the sum of these 4 compounds, which are known to contribute most significantly to PFAS levels in human serum.² Although the Food and Drug Administration (FDA) has not set specific maximum intake levels for PFAS in food, it has issued requests for information on PFAS in food, shared testing results, and even issued import alerts. The collection and evaluation of the information could be used to establish future federal guidelines or regulations.⁴ There are currently 8 PFAS (PFOA, PFOS, PFNA, PFHxS, HFPO-DA (GenX), PFBS, PFBA, and PFHxA) for which there are toxicological reference values that are used to assess potential health concerns for levels found in food. These were evaluated by the FDA, though it deemed no present human health concern based on the levels that were found in the

limited sampling of food.² However, although the FDA conducted that testing in 2021, current advances in mass spectrometry now enable the detection of a broader range of PFAS at lower concentrations, allowing for a more comprehensive and accurate assessment of PFAS in food supplies.

	PFOS	PFOA	PFNA	PFHxS	Sum of PFOS, PFOA, PFNA, and PFHxS
Eggs	1.0	0.30	0.70	0.30	1.7

Table 1. Maximum Levels (ng/g) wet weight set in EU Commission Regulation (EU) 2022/2388.²

Experimental

All standards were purchased from Wellington Laboratories. The following were used to prepare stock solutions:

Name of commercial standard	Purpose
MPFAC-HIF-ES	Extracted internal standard
MPFAC-HIF-IS	Non-extracted internal standard
PFAC-mix	Native standard mix
M2 10_2 FTS	Extracted Internal standard

Table 2. Native and isotope labeled standards used for analysis.

A native PFAS mix stock solution (500 ng/L of each analyte) was prepared in methanol and was used for serial dilutions. An extraction internal standard (EIS) solution was prepared in methanol and was used to spike egg samples prior to extraction. A mix of EIS and non-extracted internal standards (NIS) was prepared in a solution of ammonium hydroxide, water, acetic acid, and methanol, which served as diluent for the calibration curve. Lastly, the non-extracted internal standard (HIF-IS) was used to spike each eluted sample after extraction, prior to LC-

MS/MS analysis.

A 10-point solvent calibration curve in the range of 0.01 to 100 ng/mL was prepared and used for sample analysis. PFAS compound concentrations can be seen in Appendix Table 1.

Concentrations were reported in ng/g, with an EIS-spiked blank sample that was run each day subtracted from the egg samples prior to reporting values for an accurate representation of PFAS concentration (daily method blank correction). Because there is not an egg representative sample that could be used as a blank, the blank samples were 2.5 g of Q-matrix™ (CEM Corporation) that were spiked with EIS. Percent recoveries were calculated by subtracting the concentration in un-spiked eggs from that in native-spiked eggs, dividing the expected spike concentration, and multiplying 100%.

Sample Preparation

All native spiked egg samples and blanks were prepared in duplicate, and all other egg samples were prepared in triplicate. The LC-MS/MS acquisitions were also run in triplicate. Sample preparation was performed by extraction using an automated pressurized fluid extraction system (CEM EDGE PFAS®) followed by SPE on the PromoChrom Technologies SPE-03 Gen 4 Automated SPE System.

1. Add 2.5 g CEM eCleanUP Hydra and 2 g homogenous egg white and yolk mixture to assembled Q-Cup containing a Q-Disc PFAS stack
2. Spike samples respectively with EIS, Native PFAS, or leave un-spiked
Sample types: Native-spiked eggs, native-free eggs, method blanks, system blanks
3. Place Q-Cups in rack with 50 mL polypropylene conical collection tubes, and slide rack into place in the CEM EDGE PFAS™
Start method for egg extraction: 1st cycle 5 minutes with 0.02 M NaOH and 2nd cycle with 5 minutes 0.02 M NaOH
4. Concentrate collection to 2 mL (40 °C bath with nitrogen)
Reconstitute to 50 mL with LC/MS grade reagent water, vortex, check pH ≤6
5. Load 50 mL sample on PromoChrom SPE-03 with Oasis™ GCB/WAX for PFAS Cartridges (p/n: 186011112) collecting in 15 mL polypropylene collection tubes
Run SPE method following method detailed in EPA 1633 with 5 mL elute
6. To 5 mL eluted sample: add 25 µL acetic acid, spike with NIS, vortex

7. Aliquot 500 µL of sample in polypropylene vials and load on Waters Xevo™ TQ Absolute Mass Spectrometer for analysis

The dense, fatty, and viscous nature of the egg matrix makes extraction difficult. Low eCleanUP amounts (≤ 0.5 g) caused system errors and blockages, while higher loading improved dispersion, solvent flow, and sample stability. Ultimately, 2.5 g eCleanUP was chosen to ensure consistent performance.

Automated Method for PFAS in a 2 g Sample of Mixed Egg Yolk and White

Cycle	Solvent	Top add (mL)	Temp (°C)	Hold time (min)	Rinse (mL)
1	0.02 M NaOH in MeOH	5	65	3:00	0
2	0.02 M NaOH in MeOH	5	65	3:00	0
Wash	0.03% NH ₄ OH in MeOH	15	55	0:15	15
Wash	0.03% NH ₄ OH in MeOH	15	–	–	15

Table 3. CEM EDGE PFAS Method for extraction of PFAS in whole egg matrix.

LC Conditions

UPLC:	ACQUITY™ UPLC™ I-Class PLUS System with PFAS Analysis Kit
Mobile phase A:	2 mM Ammonium Acetate in Water
Mobile phase B:	2 mM Ammonium Acetate in Acetonitrile
Column(s):	Analytical column: ACQUITY Premier BEH™ C ₁₈ Column 2.1 x 50 mm, 1.7 µm (p/n: 186009452) Isolator column: Atlantis™ Premier BEH C ₁₈ AX 2.1 x 50 mm, 5.0 µm (p/n: 186010926)

Vials:	700 µL Polypropylene Screw Cap Vials (p/n: 186005219)
Column temperature:	35 °C
Sample temperature:	8 °C
Injection volume:	2 µL
Wash solvent:	50:50 MeOH: H ₂ O
Purge solvent:	10:90 MeOH: H ₂ O

Gradient Table

Time (min)	Flow rate (mL/min)	%A	%B	Curve
0	0.3	95	5	initial
0.5	0.3	75	25	6
3	0.3	50	50	6
6.5	0.3	15	85	6
7	0.3	5	95	6
8.5	0.3	5	95	6
9	0.3	95	5	6
11	0.3	95	5	6

MS Conditions

MS system:	Xevo TQ Absolute Mass Spectrometer
Ionization mode:	ESI-

Capillary voltage:	0.5 kV
Source temperature:	100 °C
Desolvation temperature:	350 °C
Desolvation flow:	900 L/hr
Cone flow:	150 L/hr
MRM transitions:	See appendix Table 2

Data Management

Software:	waters_connect™ for Quantitation
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Results and Discussion

Egg samples were analyzed as described from 2 different flocks of backyard chickens located in 2 different Massachusetts towns, as well as from organic, cage-free eggs purchased from a local grocery store, which came from a farm in New York.

Most compounds showed acceptable recoveries within the 70–130% range, as seen in Table 4, demonstrating the method’s broad application across a diverse range of PFAS analytes. The 4 compounds of regulatory focus: PFOS, PFNA, PFOA, and PFHxS all fell within this range, supporting the robustness of the method for compounds under regulatory control. Notably, the complex whole egg matrix did not significantly compromise the recoveries for most analytes, underscoring the effectiveness of the automated extraction and SPE workflow in handling challenging food matrices.

Compound	Backyard eggs % Recovery	Grocery eggs % Recovery
PFBA	112	99
PFPeA	108	96
PFHxA	117	95
PFHpA	110	99
PFOA	97	106
PFNA	104	98
PFDA	92	85
PFUnDA	112	100
PFDoDA	117	103
PFTriDA	137	122
PFTreDA	112	107
PFBS	121	109
PFPeS	117	109
PFHxS	103	100
PFHpS	118	115
PFOS	119	96
PFNS	96	88
PFDS	79	77
PFDoDS	48	56
PFEESA	117	103
PFMPA	101	92
PFMBA	114	101
GenX	111	104
ADONA	189	172
FOSA	107	91
NMeFOSA	105	99
NEtFOSA	121	107
N-MeFOSAA	109	99
N-EtFOSAA	114	99
NMeFOSE	107	99
NEtFOSE	102	95
NFDHA	64	56
9Cl-PF3ONS	115	101
11Cl-PF3OUdS	88	84
4:2 FTS	126	122
6:2 FTS	100	77
8:2 FTS	103	92
3:3 FTCA	98	86
5:3 FTCA	148	121
7:3 FTCA	124	111
PFTreDA	112	107
PFTriDA	137	122
PFUnDA	112	100

Table 4. Percent recovery of each PFAS spiked into both

backyard and grocery eggs.

The heat map found in Table 5 uses color intensity to indicate PFAS concentration levels, with warmer colors representing higher values in ng/g, and ND indicating compounds not detected after blank correction. Of the 45 compounds analyzed, 24 were detected in at least one sample. PFBA, PFPeA, and PFOS were among the most abundant compounds, with PFOS reaching levels that approached or exceeded EU regulatory thresholds in some backyard eggs. One backyard was the site of a historical house and fuel fire incident where aqueous film-forming foam (AFFF), a documented source of PFOS contamination,⁷ was likely deployed. This legacy contamination may account for the elevated PFOS concentrations observed in the eggs from chickens raised at that location. In general, backyard eggs contained a greater number of PFAS at higher concentrations than store-bought eggs, likely due to increased exposure through roaming, diet, water, bedding, etc. Precursors like FOSA and FTS were only detected in grocery store eggs, suggesting legacy contamination from older food or packaging sources. Other precursors such as NMeFOSE and emerging PFAS like ADONA and GenX were not widely detected. Backyard eggs also exhibited greater variability in PFAS concentrations, further underscoring the influence of environmental factors on PFAS exposure in non-commercial settings.

Compound	Backyard 1 eggs [ng/g]	Backyard 2 eggs [ng/g]	Grocery store organic eggs [ng/g]
PFBA	0.70	1.02	0.49
PFPeA	0.09	0.24	0.08
PFHxA	0.06	0.09	0.04
PFHpA	0.10	ND	0.10
PFOA	0.11	ND	ND
PFNA	0.11	ND	ND
PFDA	0.13	0.14	0.03
PFUnDA	0.25	0.09	0.07
PFDoDA	0.45	0.07	0.05
PFTriDA	0.55	0.12	ND
PFTreDA	0.58	0.09	ND
PFBS	ND	ND	ND
PFPeS	ND	ND	ND
PFHxS	0.07	0.03	ND
PFHpS	ND	ND	ND
PFOS	1.91	0.39	ND
PFNS	ND	ND	ND
PFDS	ND	ND	ND
PFDoDS	ND	ND	ND
PFEESA	ND	ND	ND
PFMPA	ND	ND	ND
PFMBA	ND	ND	ND
GenX	ND	ND	ND
ADONA	ND	ND	ND
FOSA	ND	ND	0.09
NMeFOSA	0.02	ND	ND
NEtFOSA	ND	ND	ND
N-MeFOSAA	ND	ND	ND
N-EtFOSAA	0.01	ND	ND
NMeFOSE	0.13	ND	0.10
NEtFOSE	ND	ND	ND
NFDHA	0.04	ND	0.04
9Cl-PF3ONS	ND	ND	ND
11Cl-PF3OUdS	ND	ND	ND
4:2 FTS	ND	ND	ND
6:2 FTS	ND	ND	0.64
8:2 FTS	ND	ND	ND
3:3 FTCA	0.33	0.15	0.24
5:3 FTCA	ND	ND	ND
7:3 FTCA	ND	ND	ND
PFTreDS	ND	ND	ND
PFTreDA	0.58	0.09	ND
PFTriDA	0.55	0.12	ND
PFUnDA	0.25	0.09	0.07
PFUnDS	ND	ND	ND

Table 5. Concentrations in ng/g of PFAS detected in

backyard and grocery store eggs.

The figures below demonstrate the presence of the 4 compounds regulated by the EU and the 8 compounds that have been evaluated in food by the FDA in the backyard and grocery store eggs evaluated. Of the 4 PFAS regulated by the EU, grocery store eggs had no-detect levels, backyard 1 had detectable levels of all 4, with PFOS above the regulatory limit, and backyard 2 had only PFHxS and PFOS detected, both below the regulatory limit.

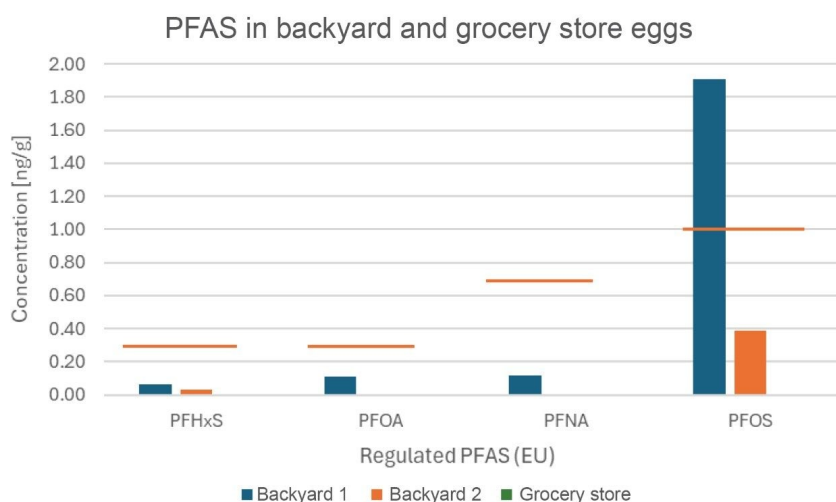


Figure 2. Four EU regulated PFAS compounds concentrations in eggs. Orange line represents maximum ng/g for PFAS in eggs.

The FDA has evaluated the four PFAS regulated by the EU, in addition to HFPO-DA (GenX), PFBS, PFBA, and PFHxS. HFPO-DA and PFBS were not detected in the grocery store or backyard eggs. While the FDA does not have regulatory limits, it was noted all 3 sources contained PFBA and PFHxA, with backyard 2 having higher levels of both compounds than backyard 1 or the grocery store eggs.

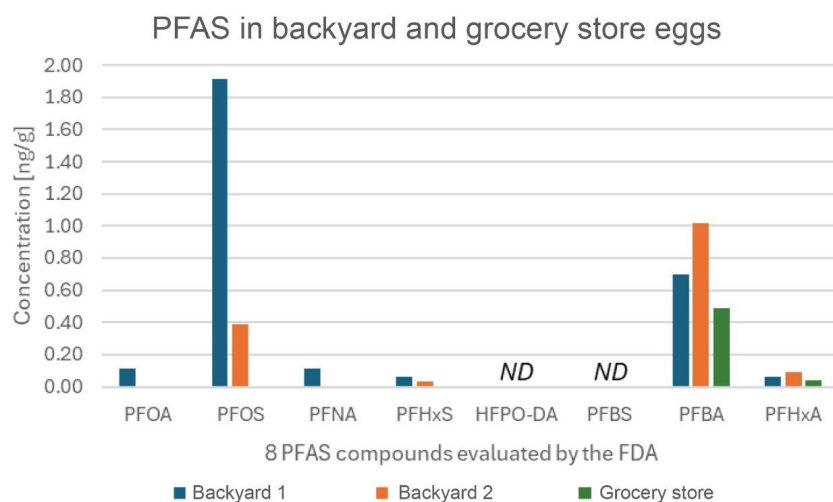


Figure 3. Eight PFAS compounds evaluated in food by the FDA. No regulatory limits have been set as of the time of this publication (May 2025).

Conclusion

The CEM EDGE PFAS and PromoChrom SPE-03 Systems, combined with the dual phase Oasis GCB/WAX for PFAS Cartridges, enabled efficient and reproducible extraction of PFAS from the challenging matrix of whole egg. Automation of the sample preparation and SPE steps delivered consistency across replicates and reduced overall method time. The workflow enabled the evaluation of PFAS compounds actively regulated by the EU. This method demonstrates that even difficult food matrices like whole egg can be prepared easily and reliably for PFAS analysis, supporting broader applications in food testing. Analysis revealed that PFOS and other PFAS were consistently higher in backyard chicken eggs compared to grocery store eggs, likely due to greater and differing environmental exposure from increased roaming space and dietary variation. PFAS precursors like FOSA were higher in grocery store eggs, likely due to potential legacy exposure. These findings highlight both the robustness of the automated method and the importance of monitoring PFAS contamination in non-commercial food sources. As awareness and monitoring of PFAS in food grows, utilization of this method offers a robust, high-confidence solution with limited user interaction — ideal for researchers and laboratories seeking to

expand testing capabilities that not only meet but outperform regulated methods, while also future-proofing workflows for emerging PFAS compounds.

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Appendix

Compound	Cal 1 (ng/mL)	Cal 2 (ng/mL)	Cal 3 (ng/mL)	Cal 4 (ng/mL)	Cal 5 (ng/mL)	Cal 6 (ng/mL)	Cal 7 (ng/mL)	Cal 8 (ng/mL)	Cal 9 (ng/mL)	Cal 10 (ng/mL)
PFBA	0.04	0.12	0.20	0.40	1.00	2.00	4.0	10.0	20.0	40.0
PFPeA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
PFHxA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFHpA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFOA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFNA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFDA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFUnDA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFDoDA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFTriDA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFTreDA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFBS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFPeS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFHxS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFHpS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFOS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFNS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFDS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFDoDS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
GenX	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
ADONA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
9ClPF3ONS	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
11ClPF3OUdS	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
4_2 FTS	0.04	0.12	0.20	0.40	1.00	2.00	4.0	10.0	20.0	40.0
6_2 FTS	0.04	0.12	0.20	0.40	1.00	2.00	4.0	10.0	20.0	40.0
8_2 FTS	0.04	0.12	0.20	0.40	1.00	2.00	4.0	10.0	20.0	40.0
FOSA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
NMeFOSA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
NEtFOSA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
NMeFOSAA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
NEtFOSAA	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
NMeFOSE	0.10	0.30	0.50	1.00	2.50	5.00	10.0	25.0	50.0	100.0
NEtFOSE	0.10	0.30	0.50	1.00	2.50	5.00	10.0	25.0	50.0	100.0
3:3 FTCA	0.04	0.12	0.20	0.40	1.00	2.00	4.0	10.0	20.0	40.0
5:3 FTCA	0.20	0.60	1.00	2.00	5.00	10.0	20.0	50.0	100.0	200.0
7:3 FTCA	0.20	0.60	1.00	2.00	5.00	10.0	20.0	50.0	100.0	200.0
PFMPA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
PFMBA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
PFEESA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
NFDHA	0.02	0.06	0.10	0.20	0.50	1.00	2.0	5.0	10.0	20.0
PFTnDS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
PFUnDS	0.01	0.03	0.05	0.10	0.25	0.50	1.0	2.5	5.0	10.0
M2_10_2 FTS	0.10	0.30	0.50	1.00	2.50	5.00	10.0	25.0	50.0	100.0
M4 PFBA	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0
M5_PFPeA	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M5_PFHxA	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M4_PFHpA	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M8_PFOA	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M9_PFNA	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M6_PFDA	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M7_PFUnDA	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M_PFDODA	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M2_PFTreDA	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M3_PFBs	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M3_PFHxS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M8_PFOS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M2_42FTS	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M2_62FTS	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M2_B2FTS	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M8_FOSA	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M3_GenX	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0	2.0
D3_NMeFOSAA	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
D5_NEtFOSAA	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
dNMeFOSA	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
dNEtFOSA	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
d7 NMeFOSE	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0
d9 NEtFOSE	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0	5.0
M3 PFBA_NIS	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0	1.0
M2 PFHxA_NIS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M4 PFOA_NIS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M5 PFNA_NIS	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
M2 PFDA_NIS	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25
18O2 PFHxS_NIS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50
M4 PFOS_NIS	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50	0.50

Appendix Table 1. Calibration curve concentrations.

Compound	PFAS group	Internal standard	CV	CE	Expected RT	Precursor mass (m/z)	Product mass (m/z)	Product mass 2 (m/z)
11Cl-PF3OUdS	ether	M8 PFOS	30	30	5.91	630.9	450.80 (Quan)	83
3:3 FTCA	carboxylate	M5 PFPeA	5	40, 10	2.14	241	176.90 (Quan)	116.9
4:2 FTS	precursor	M2 4:2 FTS	15	35, 20	2.66	326.9	307.00 (Quan)	81.1
5:3 FTCA	carboxylate	M5 PFHxA	5	25, 10	2.99	340.9	237	216.90 (Quan)
6:2 FTS	precursor	M2 6:2 FTS	15	30, 25	3.56	426.9	407	80.80 (Quan)
7:3 FTCA	carboxylate	M5 PFHxA	10	22, 17	4.42	440.9	337	316.90 (Quan)
8:2 FTS	precursor	M2 8:2 FTS	15	35, 30	4.36	526.9	506.8	80.80 (Quan)
9Cl-PF3ONS	ether	M8 PFOS	15	25	5.08	530.9	350.90 (Quan)	83
ADONA	ether	M3 GenX	10	25, 10	3.45	376.9	85	251.00 (Quan)
FBSA (PFBS)	precursor	M8 FOSA	15	30	4.69	297.9	118.9	78.00 (Quan)
FOSA	precursor	M8 FOSA	40	30	5.95	497.9	78.20 (Quan)	
GenX	ether	M3 GenX	5	35, 7	2.99	285	169.00 (Quan)	119
N-EtFOSAA	precursor	D5 N-EtFOSAA	15	20	4.82	584	525.9	418.80 (Quan)
N-MeFOSAA	precursor	D3 N-MeFOSAA	35	25, 20	4.62	569.9	418.90 (Quan)	219.1
NEtFOSA	precursor	dNEtFOSA	5	25	7.07	525.9	168.90 (Quan)	218.9
NEtFOSE	precursor	d9 NEtFOSE	15	15	6.93	630	59.00 (Quan)	
NFDHA	precursor	M5 PFHxA	5	10	2.74	295	200.90 (Quan)	84.9
NMeFOSA	precursor	dNMeFOSA	15	15	6.77	511.9	218.9	168.90 (Quan)
NMeFOSE	precursor	d7 NMeFOSE	15	15	6.62	616	59.00 (Quan)	
PFBA	carboxylate	M4 PFBA	10	20	2	212.9	169.00 (Quan)	19
PFBS	sulfonate	M3 PFBS	15	30	2.92	298.9	80.10 (Quan)	99.1
PFDA	carboxylate	M6 PFDA	14, 15	15, 9	4.47	512.9	468.90 (Quan)	219
PFDoDA	carboxylate	M PFDoDA	30	25, 10	5.28	612.9	568.90 (Quan)	169
PFDoDS	sulfonate	M8 PFOS	40	55	6.3	699.1	80.00 (Quan)	99
PFDS	sulfonate	M8 PFOS	25	40	5.51	598.9	80.20 (Quan)	99.1
PFEESA	precursor	M5 PFHxA	15	20	3.11	314.9	134.90 (Quan)	82.9
PFHpA	carboxylate	M4 PFHpA	15	15, 10	3.22	362.9	319.00 (Quan)	169
PFHpS	sulfonate	M8 PFOS	15	35	4.24	448.9	80.20 (Quan)	99.1
PFHxA	carboxylate	M5 PFHxA	5	20, 10	2.8	312.9	269.00 (Quan)	119
PFHxS	sulfonate	M3 PFHxS	10	35, 30	3.81	398.9	80.10 (Quan)	99.1
PFMBA	carboxylate	M5 PFPeA	10	10	2.49	278.9	84.90 (Quan)	
PFMPA	carboxylate/ether	M5 PFPeA	23	10	2.13	228.9	84.90 (Quan)	
PFNA	carboxylate	M9 PFNA	10	15, 10	4.07	462.9	418.90 (Quan)	219
PFNS	sulfonate	M8 PFOS	20	40	5.11	548.9	80.20 (Quan)	99.2
PFOA	carboxylate	M8 PFOA	10	15, 10	3.66	412.9	369.00 (Quan)	169
PFODA	carboxylate	M2 PFTreDA	30	25, 10	7.79	912.9	868.90 (Quan)	169.2
PFOS	sulfonate	M8 PFOS	15	40	4.69	498.9	80.20 (Quan)	99.1
PFPeA	carboxylate	M5 PFPeA	10	20, 5	2.36	262.9	219.00 (Quan)	19
PFPeS	sulfonate	M3 PFHxS	10	30	3.43	348.9	80.10 (Quan)	99.1
PFTreDA	carboxylate	M2 PFTreDA	10	25, 15	6.09	712.9	668.90 (Quan)	169
PFTriDA	carboxylate	M2 PFTreDA	5	30, 10	5.7	662.9	618.90 (Quan)	169
PFUnDA	carboxylate	M7 PFUnDA	25	20, 10	4.88	562.9	518.90 (Quan)	269
PFTTrDS	sulfonate	M2 10:2 FTS	40	54, 64	6.8	749.1	80.00 (Quan)	99
PFUnDS	sulfonate	M2 10:2 FTS	40	52, 48	6.03	649.1	80.00 (Quan)	99

Appendix Table 2. MS method conditions for PFAS included in method. “(Quan)” indicates MRM used as quan ion.

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