

## Performance Characteristics of the MassTrak Vitamin D Solution

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Waters Corporation

For *in vitro* diagnostic use. Not available in all countries.

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### Abstract

The Waters MassTrak Vitamin D Solution is designed for the quantitative determination of serum or plasma 25-hydroxyvitamin D<sub>3</sub> (25OHD<sub>3</sub>) and 25-hydroxyvitamin D<sub>2</sub> (25OHD<sub>2</sub>), which in combination provide the total 25-hydroxyvitamin D concentration as an aid in the assessment of vitamin D sufficiency.

### Benefits

- Independent measurement of 25OHD<sub>2</sub> and 25OHD<sub>3</sub>
  - Aids laboratory compliance to ISO 15189; Calibrators and quality control sets are traceable to NIST Standard Reference Material (SRM) 2972
  - Streamlined workflow facilitates high sample throughput utilizing automation in a multi-well plate format while enabling sample tracking from the sample bar code to data import into MassLynx Software
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## Introduction

Assessment of vitamin D status is routinely performed to assess body stores of vitamin D and as an aid to diagnose; vitamin D deficiency or intoxication, intestinal mal-absorption, and to monitor adherence to therapy, and the therapeutic response in patients during treatment for vitamin D related disorders.

Vitamin D<sub>3</sub> (cholecalciferol) is produced in the skin when the UV-B portion of sunlight converts 7-dehydrocholesterol to previtamin D<sub>3</sub>, which undergoes thermal isomerization to form vitamin D<sub>3</sub>. Latitude, season, aging, sunscreen use, clothing, and skin pigmentation influence production of vitamin D<sub>3</sub> in the skin. Vitamin D<sub>2</sub> (ergocalciferol) is produced through UV irradiation of ergosterol, which is found naturally in certain plankton, yeasts, and fungi and is used as a vitamin D dietary supplement in many countries<sup>1</sup>. Both vitamin D<sub>2</sub> and vitamin D<sub>3</sub> are metabolized, primarily in the liver, to 25-hydroxyvitamin D (25OHD). This accepted indicator of vitamin D status (Total 25OHD, which is the sum of 25OHD<sub>2</sub> and 25OHD<sub>3</sub>) which has been a challenge to measure accurately because the antibodies used in many immunoassays do not have 100% co-specificity for both 25OHD<sub>2</sub> and 25OHD<sub>3</sub>.

The Waters MassTrak Vitamin D Solution is designed for the quantitative determination of serum or plasma 25-hydroxyvitamin D<sub>3</sub> (25OHD<sub>3</sub>) and 25-hydroxyvitamin D<sub>2</sub> (25OHD<sub>2</sub>), which in combination provide the total 25-hydroxyvitamin D concentration as an aid in the assessment of vitamin D sufficiency.

The MassTrak Vitamin D Kit, a component of the MassTrak Vitamin D Solution is validated for use with the Waters ACQUITY UPLC I-Class/Xevo TQD IVD System and the sample preparation has been validated using the Tecan Freedom 100/4 EVO Offline Automated Liquid Handling System.



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Figure 1. The Waters ACQUITY UPLC I-Class/Xevo TQD IVD System and MassTrak Vitamin D Kit.

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## Experimental

### LC conditions

System:	ACQUITY UPLC I-Class with FTN
Needle:	30 $\mu$ L
Column:	MassTrak Vitamin D BEH Phenyl 130 Å, 1.7 $\mu$ m, and 2.1 mm x 50 mm IVD (p/n: 186008647IVD)

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Pre-Column filter:	Column In-Line Filter Kit
Mobile phase A:	Aqueous 2 mM ammonium acetate with 0.1% formic acid
Mobile phase B:	Methanol with 2 mM ammonium acetate with 0.1% formic acid
Column temp:	35 °C
Injection volume:	20 µL
Flow rate:	0.45 mL/min
Gradient:	See Table 1
Run time:	4.2 minutes

## Data management

MassLynx v4.1 SCN 918IVD (p/n: 667005037IVD) with TargetLynx IVD v4.1 Application Manager (p/n: 667002671IVD)

Tecan File Converter Software v2.0 (p/n: 667004831)

## MS conditions

System:	Xevo TQD
Resolution:	MS1 (0.7 FWHM)
	MS2 (0.85 FWHM)

Acquisition mode: Multiple Reaction Monitoring (MRM) (see Table 2 for details)

Polarity: ESI positive

Capillary: 0.80 kV

Source temp.: 120 °C

Desolvation temp.: 400 °C

Refer to the Directions for Use (p/n: [715004830IVD < https://www.waters.com/webassets/cms/support/docs/715004830ivdrh.pdf>](https://www.waters.com/webassets/cms/support/docs/715004830ivdrh.pdf) ) and the System Operator' s Guide (p/n: [715004831IVD < https://www.waters.com/webassets/cms/support/docs/715004831ivdrg.pdf>](https://www.waters.com/webassets/cms/support/docs/715004831ivdrg.pdf) )

## Method conditions

Time (min)	Flow rate (mL/min)	%A	%B	Curve
Initial	0.450	35	65	Initial
2.50	0.450	20	80	6
2.70	0.450	2	98	11
3.50	0.450	35	65	11

Table 1. Gradient table for the separation of 25OHD<sub>2</sub> and 25OHD<sub>3</sub>.

Analyte	Precursor ion (m/z)	Product ion (m/z)	Cone voltage (V)	Collision energy (kV)
25OHD <sub>3</sub> (Quan)	401.3	159.1	24	24
25OHD <sub>3</sub> (Qual)	401.3	365.3	24	10
[ <sup>2</sup> H <sub>3</sub> ]-25OHD <sub>3</sub>	404.3	162.1	24	24
25OHD <sub>2</sub> (Quan)	413.3	355.3	24	10
25OHD <sub>2</sub> (Qual)	413.3	83.1	24	24
[ <sup>2</sup> H <sub>3</sub> ]-25OHD <sub>2</sub>	416.3	358.3	24	10

Table 2. MRM parameters of 25OHD<sub>2</sub> and 25OHD<sub>3</sub> quantifier ions, qualifier ions, and their internal standards.

## Sample Preparation

The sample preparation process is automated for the MassTrak Vitamin D Solution and is performed using the Tecan Freedom EVO 100/4 Offline Automated Liquid Handling System.\* The sample preparation consists of a series of steps that selectively extract 25OHD<sub>2</sub> and 25OHD<sub>3</sub> from the serum or plasma sample matrix ready for quantitative analysis by LC-MS/MS in less than two hours for 96 samples. A Tecan File Converter program is run on the Tecan workstation PC during sample preparation to collect the sample sequence information from the sample barcodes, and this is used to create an accurate sample list in the MassLynx Software.

A fixed quantity of internal standard solution is added to 150 µL of each test sample (calibrator, quality control, and patient sample). This is followed by the addition of zinc sulphate solution, which aids the release of the 25OHD from vitamin D-binding protein and denatures proteins to facilitate their precipitation. Methanol is added to complete the denaturing and precipitation process of the proteins. Samples are centrifuged to separate the protein precipitate from the 25OHD and the supernatant loaded onto an Oasis HLB µElution SPE Plate. 25OHD<sub>2</sub> and 25OHD<sub>3</sub> and their corresponding internal standards are separated from interfering compounds using specific solvent washes and eluted into 96-well plates ready for transfer to the ACQUITY UPLC Sample Manager (SM-FTN) for analysis.

## Sample analysis

The analytes are separated on a MassTrak Vitamin D BEH Phenyl IVD Column using the ACQUITY UPLC I-

Class System prior to the multiple reaction monitoring (MRM) detection and quantitation by the Xevo TQD. The data are acquired using the MassLynx Software and sample concentrations are calculated using TargetLynx Application Manager.

## Results and Discussion

### Performance Characteristics

Precision was verified in a multi-site precision evaluation conducted in three sites using a total of three systems, three operators, and a single reagent kit lot according to CLSI EP05-A3. Five serum pools (Panel 1–5) spanning the range of the assay from 22 nmol/L to 300 nmol/L for 25OHD<sub>2</sub> and 25OHD<sub>3</sub> and the Quality Controls (Low and High) were analyzed in replicates of five, for five days, at three sites, providing a reproducibility of ≤5.9%CV for 25OHD<sub>2</sub> (range 4.1–6.1%) and ≤6.8%CV for 25OHD<sub>3</sub> (range 3.9–6.8%) as shown in Tables 3 and 4 respectively.

25OHD <sub>2</sub>	Mean	Between run		Between day		Between site		Reproducibility	
	nmol/L	SD	%CV	SD	%CV	SD	%CV	SD	%CV
Panel 1	25.32	1.13	4.5	0.00	0.0	0.75	3.0	1.36	5.4
Panel 2	47.73	2.80	5.9	0.32	0.7	0.00	0.0	2.82	5.9
Panel 3	98.74	3.43	3.5	1.83	1.9	1.39	1.4	4.13	4.2
Panel 4	187.15	11.07	5.9	0.00	0.0	2.69	1.4	11.40	6.1
Panel 5	299.12	9.91	3.3	7.37	2.5	0.00	0.0	12.35	4.1
QC Low	61.48	2.42	3.9	0.79	1.3	1.97	3.2	3.21	5.2
QC High	218.21	7.53	3.5	6.72	3.1	4.23	1.9	10.95	5.0

Table 3. Multi-site precision performance for 25OHD<sub>2</sub>.

25OHD <sub>3</sub>	Mean	Between run		Between day		Between site		Reproducibility	
	nmol/L	SD	%CV	SD	%CV	SD	%CV	SD	%CV
Panel 1	24.99	1.69	6.8	0.21	0.8	0.00	0.0	1.70	6.8
Panel 2	50.85	3.14	6.2	0.00	0.0	0.53	1.0	3.19	6.3
Panel 3	104.19	3.97	3.8	1.91	1.8	0.58	0.6	4.44	4.3
Panel 4	206.55	11.43	5.5	0.00	0.0	4.82	2.4	12.45	6.0
Panel 5	311.89	8.93	2.9	8.15	2.6	0.00	0.0	12.10	3.9
QC Low	57.46	2.58	4.5	1.54	2.7	0.00	0.0	3.00	5.2
QC High	199.54	7.67	3.8	4.50	2.3	3.61	1.8	9.60	4.8

Table 4. Multi-site precision performance for 25OHD<sub>3</sub>.

The reportable range for linearity for 25OHD<sub>2</sub> and 25OHD<sub>3</sub> was demonstrated to be from 10 nmol/L to 375 nmol/L, within a range of  $\pm 10\%$  for this interval (Figure 2). Linearity was assessed in a single site study according to CLSI EP6-A.

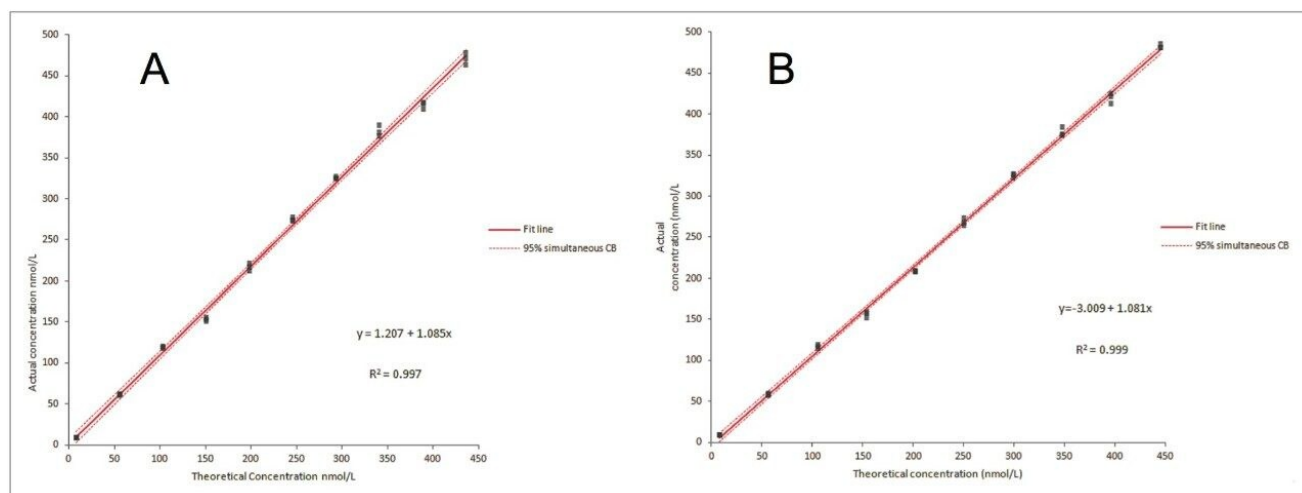


Figure 2 Linearity of 25OHD<sub>2</sub> (A) and 25OHD<sub>3</sub> (B).

Sensitivity testing was conducted in accordance with CLSI EP 17-A2 and the Limit of Detection and Limit of Quantification were 2.7 nmol/L and 7.3 nmol/L, respectively, for 25OHD<sub>3</sub>. The Limit of Detection and Limit of Quantification were 2.7 nmol/L and 5.7 nmol/L, respectively, for 25OHD<sub>2</sub>.



The carryover of the assay was determined to be 0.87 nmol/L for 25OHD<sub>2</sub> and 0.37 nmol/L for 25OHD<sub>3</sub>. The total carryover effect for 25OHD was 1.24 nmol/L which is lower than the limit of quantitation of the assay.

Interference testing was conducted, according to the procedures described in CLSI EP7-A. Both endogenous and exogenous compounds were tested. Sample collection tube preservatives were tested (potassium-EDTA, sodium citrate, sodium heparin, and lithium heparin) as well as certain vitamin D metabolites (1- $\alpha$ -(OH)vitamin D<sub>3</sub>, 1- $\alpha$ -(OH)vitamin D<sub>2</sub>, 1,25di(OH)vitamin D<sub>2</sub>, 1,25di(OH)vitamin D<sub>3</sub>, 3-epi-25(OH)vitamin D<sub>2</sub>, 3-epi-25(OH)vitamin D<sub>3</sub>, 23R,25di(OH)vitamin D<sub>3</sub>, 24R,25di(OH)vitamin D<sub>3</sub>, 24S,25di(OH)vitamin D<sub>3</sub>). The mean recovery for 25OHD<sub>2</sub> and 25OHD<sub>3</sub> was between 85%–115% in the presence of the endogenous, exogenous and vitamin D metabolites tested, with the exception of 3-epi-25OHD<sub>2</sub> and 3-epi-25OHD<sub>3</sub>. Interference was observed from 3-epi-25OHD<sub>2</sub> for 25OHD<sub>2</sub> and 3-epi-25OHD<sub>3</sub> for 25OHD<sub>3</sub>, which co-elute with the analytes of interest.

A sample dilution protocol using 1:1 proportion of sample to calibrator 0 was developed and demonstrated to provide recovery within 85–115% for out of range samples (>375 nmol/L of 25OHD<sub>2</sub> or 25OHD<sub>3</sub>).

The accuracy of the MassTrak Vitamin D Kit has been demonstrated. Waters enrolls in the VDSCP for total serum 25OHD. The program assesses bias and precision of assays relative to reference measurement procedures. The MassTrak Vitamin D assay achieved a mean bias of 0.6% from the VDSCP reference values and a mean imprecision of 4.9%, meeting the certification performance criteria. The linear regression analysis agreement between mean MassTrak Vitamin D Kit measured concentrations (n=40) and the values assigned by the reference measurement procedure was described by the equation: MassTrak Vitamin D Kit = 1.1143(VDSCP) - 7.5584, with R<sup>2</sup>= 0.9715.

Finally, metrological traceability of the MassTrak Vitamin D Kit calibrators and quality controls to NIST SRM2972 has been established aiding laboratories in their compliance to ISO 15189 ([720005886EN](#)).

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## Conclusion

The MassTrak Vitamin D Solution provides an LC-MS/MS based clinical diagnostic solution that is CE Marked to the IVDR (EU) 2017/746 for quantitative assessment of vitamin D status in human plasma or

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serum.

The MassTrak Vitamin D Solution demonstrates excellent precision and linearity across a wide range. The limit of quantitation has been determined to be 7.3 nmol/L for 25OHD<sub>3</sub> and 5.7 nmol/L for 25OHD<sub>2</sub>. The C3-epimers; 3-epi-25OHD<sub>2</sub> and 3-epi-25OHD<sub>3</sub> were demonstrated to be the only interference in the method. Therefore, in samples containing high concentrations of these C3-epimers, 25OHD<sub>2</sub> and 25OHD<sub>3</sub> can be overestimated.

In addition, the time consuming sample preparation steps have been automated to minimize operator and transcription errors in sample processing with sample tracking from the sample bar code through to reporting of the result in TargetLynx.

Metrological traceability of commercial calibrators is important to aid compliance to ISO 15189 in your laboratory. The MassTrak Vitamin D Kit calibrators and quality controls are traceable to NIST SRM2972 with uncertainty measurements provided in the Directions for Use (p/n: [715004830IVD](#) < <https://www.waters.com/webassets/cms/support/docs/715004830ivdrh.pdf> ) to aid compliance.

Intended Use: The Waters MassTrak Vitamin D Kit is for the quantitative determination of 25-hydroxyvitamin D<sub>3</sub>, 25-hydroxyvitamin D<sub>2</sub>, which in combination provide the total 25-hydroxyvitamin D in human plasma and serum using an automated liquid handling system and Waters ACQUITY UPLC I-Class/Xevo TQD IVD System. Results are to be used as an aid in the assessment of vitamin D sufficiency.

For information on availability, please contact your local sales representative.

*\*Not available from Waters, must be ordered from Tecan.*

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[MassLynx \(IVD\) Mass Spectrometry Software with TargetLynx \(IVD\)](#) <

<https://www.waters.com/waters/nav.htm?cid=134834177>>

[MassTrak Vitamin D Solution](#) <<https://www.waters.com/waters/nav.htm?cid=134905460>>

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