

Application Note

Fast Analysis of Isoflavones in Dietary Supplements - Method Transfer to the ACQUITY UPLC H-Class System

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Abstract

In this application note, we further shortened the analysis time to 8.1 min, while meeting performance requirements using an ACQUITY H-Class UPLC System. The method performance characteristics of this UPLC method are presented.

Benefits

- Fast analysis of isoflavones in 8 minutes on an ACQUITY UPLC H-Class System
- Waters ACQUITY QDa Mass Detector speeds up method development and reduces interference

Introduction

Major isoflavones that are primarily found in soy (*Glycine max*), red clover (*Trifolium pretense*), and Kudzu (*Pueraria lobata*) are daidzein, glycitein, genistein, and their respective glucoside derivatives. The structures of 12 major isoflavones and apigenin, an internal standard (IS), are shown in Figure 1. Methods for the determination of isoflavones in dietary supplements have been established.^{1,2} These standard methods are based on reversed-phase high performance liquid chromatography (RP-HPLC) and ultraviolet and visible light (UV-Vis) spectroscopy detection. Because of the structural similarity and the large number of isoflavones, the method chromatographic run times are more than 70 minutes long. Recently, we have transferred an isoflavones standard method¹ onto an ACQUITY Arc UHPLC System.³ The run time of the new method was reduced to 18 minutes per injection cycle. We also demonstrated the benefits of a small but powerful mass spectrometer, the ACQUITY QDa Mass Detector.^{4,5} With the ACQUITY QDa Detector, the time spent in the method transfer and development was greatly reduced, and the risk of interference was significantly lowered. In this application note, we further shortened the analysis time while meeting performance requirements using an ACQUITY H-Class UPLC System. The method performance characteristics of this UPLC method were presented.

System:	ACQUITY UPLC H-Class with ACQUITY PDA and ACQUITY QDa Detector (Performance)
Software:	Empower 3
Column:	CORTECS UPLC C ₁₈ 2.1 x 75 mm, 1.6 µm
Column temp.:	30 °C
Mobile phase A:	Water with 0.1% formic acid
Mobile phase B:	ACN with 0.1% formic acid
Inj vol.:	0.7 µL
Flow rate:	0.55 mL/min
Run time:	8.10 min
UV detection:	260 nm
UV resolution:	1.2 nm

Elution gradient:

	Time (min)	Flow rate (mL/min)	%A	Curve
1	Initial	0.55	90	6
2	6.48	0.55	70	6
3	6.53	0.55	10	6

4	6.84	0.55	10	6
5	6.93	0.55	90	6
6	8.1	0.55	90	6

QDa parameters

Polarity:	ES+
Capillary (kV):	0.8
Cone (V):	15
Probe temp.:	600 °C

SIR masses:

Analyte	SIR mass (Daltons)
Daidzin	417.1
Glycitin	447.0
Genistin	433.1
Daidzein	254.9
Glycitein	285.0
Genistein	270.9
Apigenin (IS)	270.9
Malonyl daidzin	503.4
Malonyl glycitin	533.1
Acetyl daidzin	459.1
Acetyl glycitin	489.0
Malonyl genistin	519.0
Acetyl genistin	475.1

Table 1. Masses of isoflavone molecular ions.

Results and Discussion

1) Method transfer using UPLC column calculator

The USP method (isoflavones powder extract)¹ was transferred to an ACQUITY UPLC H-Class System with a CORTECS UPLC C₁₈ Column (1.6 μ m, 2.1 x 75 mm). The CORTECS columns are packed with solid-core particles, which provide higher separation efficiency and lower back pressure than fully porous particles of equivalent particle size. The USP method's HPLC gradient elution program was converted to a new UPLC gradient elution program using a Waters ACQUITY UPLC Column Calculator.³ The column properties of the HPLC and the UPLC methods were entered in the Column Calculator software to generate an equivalent UPLC elution program. This UPLC elution program was then used as the starting point and was optimized in the method development phase. The mobile phase additive was changed from 0.05% phosphoric acid to 0.1% formic acid, which is a mass spectrometry (MS) friendly additive. The factory default ACQUITY QDa instrument parameters were used without any modification.

2) Method development with QDa Mass Detector

As the acetyl and the malonyl isoflavone standards are not commercially available, the peak assignment of these compounds relied on a reference material, the USP defatted powdered soy RS.¹ By comparing the UV chromatograms of the unheated and the heated reference samples to their reference chromatograms, one can assign the peak IDs to the appropriate isoflavones. However, due to the change in LC conditions (HPLC to UPLC), and the close-eluting peaks, the UV peak patterns obtained in UPLC did not perfectly match those in the HPLC reference chromatograms. This issue was easily solved by using MS Single Ion Recording (SIR) traces (see Figure 2). Table 1 lists the isoflavone molecular ion masses. The Waters ACQUITY QDa Detector selectively detected these compounds in separate SIR channels, and avoided any possible interference from co-eluting compounds. The column temperature and the flow rate were optimized using the reference material. The optimization process was relatively quick since only one injection of the reference material generated enough information (peak RT and resolution) for one particular LC condition (column temperature or flow rate). Without the ACQUITY QDa Detector, usually several injections of standard solutions would have been needed for each LC condition in the method optimization process.

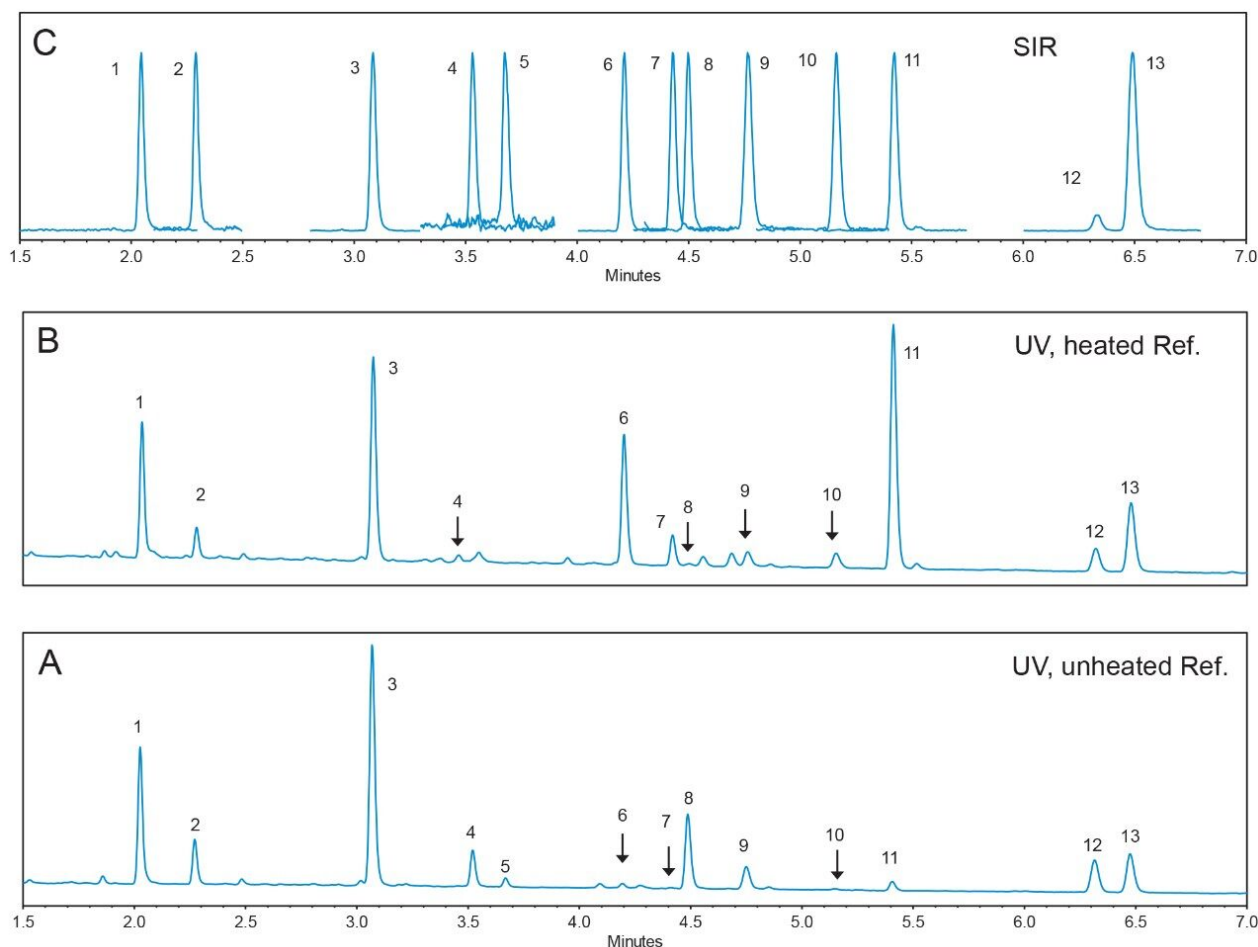


Figure 2. A) UV chromatogram of the unheated reference sample (USP defatted powdered soy RS), B) UV chromatogram of the heated reference sample, and C) the corresponding normalized SIR trace overlay for each compound. Peak ID: 1) daidzin; 2) glycitin; 3) genistin; 4) malonyl daidzin; 5) malonyl glycitin; 6) acetyl daidzin; 7) acetyl glycitin; 8) malonyl genistin; 9) daidzein; 10) glycitin; 11) acetyl genistein; 12) genistein; 13) apigenin.

3) Method performance characteristics

Table 2 shows the UV calibration results for the available standards. Apigenin was used as an IS for the calibration and the quantitation of all compounds. The square of the correlation coefficients (R^2) between the responses (peak area ratio) and the standard concentrations (ppm) for all compounds were better than 0.998. The relative standard deviation (RSD) in retention time for all compounds were less than 0.27%.

Table 3 shows the isoflavone results for the NIST 3238 SRM and a comparison to its certified and reference values. Relative difference of less than 14% was obtained for the genistin, glycitin, daidzin, genistein, and

glycitein. The accuracy for the daidzein, genistein, and glycitein was also evaluated by a spiking experiment (Table 4). Recovery of 95% to 102% were obtained for these analytes.

Analyte	RT		Equation	R ²	Range (ppm)
	Min	RSD (%)			
Daidzin	2.03	0.21	$y = 2.12 \times 10^{-1} X + 3.93 \times 10^{-3}$	0.999	0.075–10
Glycitin	2.27	0.27	$y = 2.45 \times 10^{-1} X + 4.54 \times 10^{-3}$	0.998	0.075–10
Genistin	3.07	0.21	$y = 4.56 \times 10^{-1} X + 5.30 \times 10^{-3}$	0.998	0.075–10
Daidzein	4.76	0.11	$y = 2.61 \times 10^{-1} X + 4.50 \times 10^{-3}$	0.998	0.075–10
Glycitein	5.15	0.14	$y = 4.97 \times 10^{-1} X + 5.69 \times 10^{-3}$	0.998	0.05–10
Genistein	6.32	0.09	$y = 4.88 \times 10^{-1} X + 4.87 \times 10^{-3}$	0.998	0.05–10

Table 2. Isoflavones retention times and their relative standard deviation, calibration equations, R², and linear ranges.

	NIST value (mg/kg)	Determined value (mg/kg)	Rel. difference
Genistin	12700±530	11432	-10%
Glycitin	3760±180	3669	-2%
Daidzin	13400±2400	15102	13%
Daidzein	241±5	262	9%
Genistein	108±10	93	14%
Glycitein	211±5	190	10%

Table 3. Comparison of determined isoflavone values to the certified and reference values of NIST 3238 SRM.

	Original value (mg/kg)	Spiked level (mg/kg)	Determined value (mg/kg)	Recovery (%)
Daidzein	317	499	827	102%
Genistein	159	499	634	95%
Glycitein	165	499	665	100%

Table 4. Recovery results from a spiking experiment.

4) Analysis of isoflavones in dietary supplements

The isoflavone contents in three isoflavone dietary supplement samples were measured by this 8-minute

UPLC-UV method. The sample forms included tablets, capsules, and powder. The USP calibration and quantitation protocols¹ were followed in the data processing. The conversion factors for the acetyl and malonyl derivatives that were specified in the USP HPLC-UV method were used in this analysis. Table 5 shows the determined individual and total isoflavone contents, as well as the total isoflavone contents on labels. For an easy comparison, the label values were converted to the same concentration unit (mg/kg). Two of the three samples (C and D) showed good agreement between the determined values and their label values, while one sample (B) had much less measured total isoflavone content than its label value. The reason for this low total isoflavone content in sample B compared to its label claim is unknown.

Sample (mg/kg)	B		C		D	
	Mean	%RSD	Mean	%RSD	Mean	%RSD
Daidzin	3095	1.0	12837	0.2	310	4.3
Glycitin	617	2.0	4792	0.2	0	
Genistin	271	3.2	1766	0.6	664	1.1
Malonyl daidzin	97	7.8	655	2.0	0	
Acetyl daidzin	1634	1.7	7391	0.3	0	
Acetyl glycitin	390	1.5	2202	0.04	0	
Malonyl genistin	0		0		221	1.2
Daidzein	6418	0.8	407	0.7	0	
Glycitein	0		179	1.4	0	
Acetyl genistin	162	2.1	1042	0.4	0	
Genistein	0		0		0	
Total isoflavones	12,684		31,270		1,195	
Label claim value	>31,250		>25,000		867–2,600	

Table 5. Isoflavone content in dietary supplements and their label claim values.

5) Benefits of UPLC and MS

Figure 3 is a comparison of the UPLC and the UHPLC chromatograms of the same reference material (USP defatted soy RS). The total run time in the UPLC method was 8.1 min, which was less than a half of the total run time in the UHPLC method (18 min). The UPLC method can provide twice the sample analysis throughput than the UHPLC method. The solvent cost for UPLC was less than a quarter of that for UHPLC since the UPLC flow rate was about half that of UHPLC. Both of the UHPLC and the UPLC methods were much faster than the existing USP method, which is a 74 min HPLC method.

The MS detector was extremely useful during method transfer and development and in complex sample analyses. Compared to typical UV/Vis detection, the ACQUITY QDa Detector provided excellent detection selectivity. This high selectivity offered high confidence in peak assignment, which was extremely useful

when pure standards were not available and interference was common.

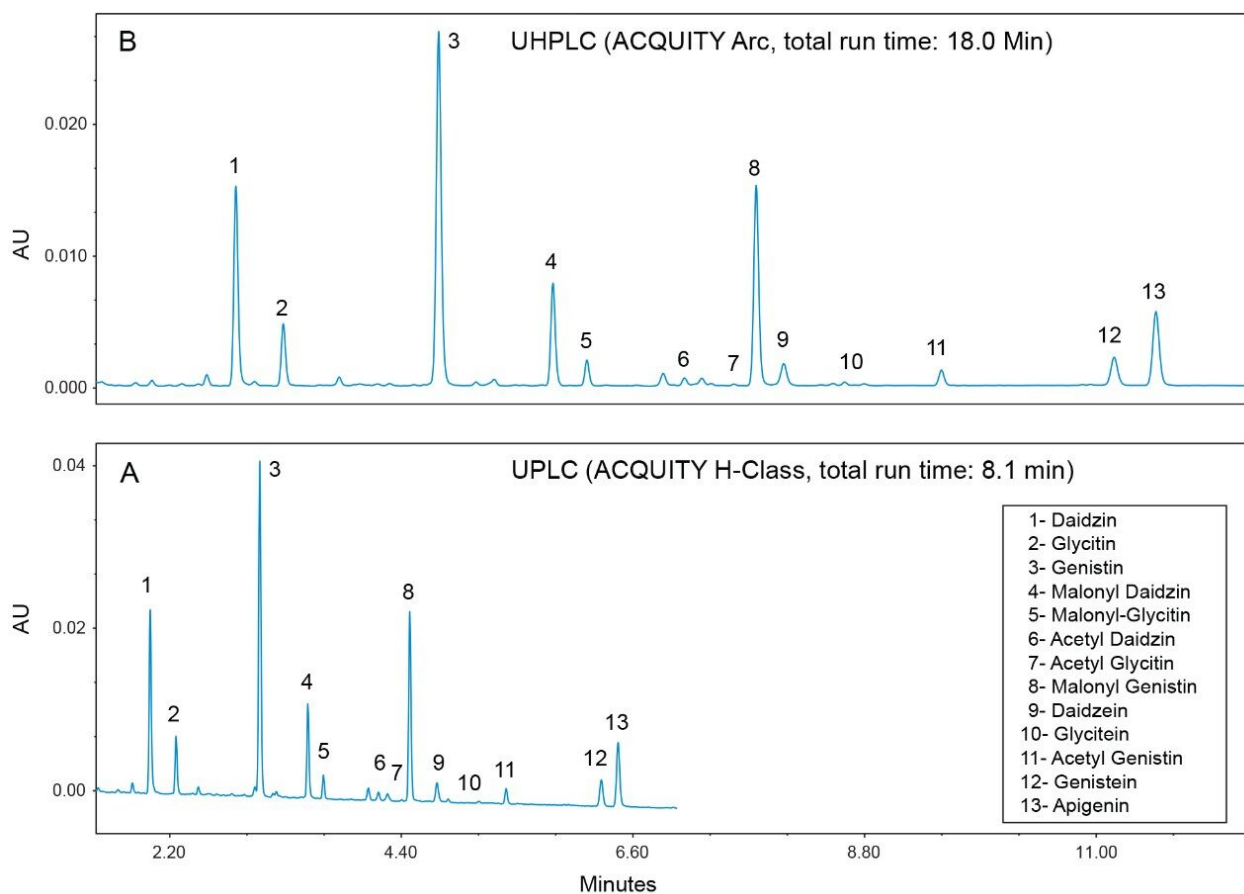


Figure 3. Comparison of (A) UPLC and (B) UHPLC chromatograms of the reference material (unheated).

Conclusion

The USP isoflavone method was successfully transferred to an ACQUITY UPLC H-Class System with an ACQUITY PDA Detector. The total analysis time per injection was 8.1 minutes, which was less than the half of the UHPLC method (18 min),⁵ and significantly shorter than the HPLC methods (74 minute).^{1,2} The UPLC method provides 9 times higher analysis throughput, and more than 90% cost savings in solvents than the HPLC methods. ACQUITY QDa Mass Detector offered excellent detection selectivity, which was a great asset in the method transfer and development, as well as in the isoflavone analysis for unknown and challenging samples where potential interference from co-eluting compounds can be high. Analysis of

isoflavones in three dietary supplements showed compliance with the label claims for two of the samples. One sample showed low total isoflavone content comparing to its label claim.

References

1. USP Monograph. Powdered Soy Isoflavones Extract, USP39–NF34 S1 [6841], The United States Pharmacopeial Convention.
2. AOAC Official Method 2008.03 Total soy isoflavones in dietary supplements, supplement ingredients, and soy foods HPLC with UV detection First Action 2008, AOAC International 2008.
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4. Jinchuan Yang, Mark Benvenuti, Gareth Cleland, Fast Analysis of Isoflavones – Benefits of Mass Detection in Method Transfer and Sample Analysis, Waters application note, [to be published], 2016.
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