

Analysis of Labeled and Non-Labeled Proteomic Data Using Progenesis QI for Proteomics

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Abstract

This application note demonstrates the ability of Progenesis QI for proteomics to achieve qualitative and quantitative results for both labeled and label free quantitation workflows.

Benefits

Data acquired by means of data-dependent acquisition and data-independent acquisition strategies show consistency and precision in their reported quantitation values, independent of the quantitation methodology adopted.

Introduction

LC-MS is routinely applied for the qualitative and quantitative analysis of complex proteomes, to characterize biological processes, and understand disease states. Experiments can readily generate large, complex data sets

in effect making the analysis and interpretation of results rate-determining steps. This lag factor has created a demand for improved data-analysis systems including efficient and accurate data-compression routines, intuitive software interfaces with menu-guided workflows, flexible experimental designs without sample-number restrictions, consistent peak detection for improved accuracy and precision, complete data matrices without missing values for reliable statistics, and the ability to analyze fractionated samples. These features, core to the Progenesis Q1 for proteomics solution, will be illustrated for both the identification and quantification of isotopically-labeled and label-free proteomic data sets.

Experimental

Samples

- Cytosolic *Escherichia coli* (*E. coli*) tryptic protein digest spiked with bovine serum albumin (BSA), alcohol dehydrogenase (ADH), enolase, and glycogen phosphorylase B digest standards
- Tryptic digest dimethyl-labeled HL60 human B cells (University of Leiden)
- Sigma-Aldrich UPS1 standard (25, 2.5, and 0.125 fmol) spiked into a cell lysate of *Saccharomyces cerevisiae* (yeast)

LC conditions

LC system:	nanoACQUITY UPLC
Column:	5 μm Symmetry C ₁₈ , 180 μm x 20 mm 2G Trap Column and a 1.8 μm HSS T3 C ₁₈ , 75 μm x 150 mm NanoEase Analytical Column
Column temperature:	35 °C
Flow rate:	300 nL/min

Mobile phase:	water (0.1% formic acid) (A) and acetonitrile (0.1% formic acid) (B)
Gradient:	3% to 40% B in 90 min
Injection volume:	1 μ L

MS conditions

MS system:	SYNAPT G2- <i>Si</i>
Ionization mode:	ESI (+) at 3.2 kV
Cone voltage:	30 V
Acquisition range:	50 <i>m/z</i> to 2000 <i>m/z</i> (DDA and HDMS ^E)
Acquisition rate:	0.2 s (DDA) or 0.5 s (HDMS ^E)
Collision energy (HDMS ^E):	5 eV (low energy) and from 19 eV to 45 eV (elevated energy)
Collision energy (DDA):	<i>m/z</i> -dependent ramp

Data management

Progenesis Q1 for proteomics v2.0 (Nonlinear Dynamics, Newcastle Upon Tyne, UK)

ProteinLynx Global SERVER v3.0.2 (Waters Corporation, Milford, MA)

ProteoLabels v1.0 (University of Liverpool, UK)

Mascot v2.5 (Matrix Science, London, UK)

Results and Discussion

As illustrated in Figure 1, peak detection is conducted first [1]. To assess the precision of peak detection, the separate data and detected peaks and features from six technical LC-IM-DIA-MS replicates of an *E. coli* digest were compared, providing an average of $28,793 \pm 458$ detected features.

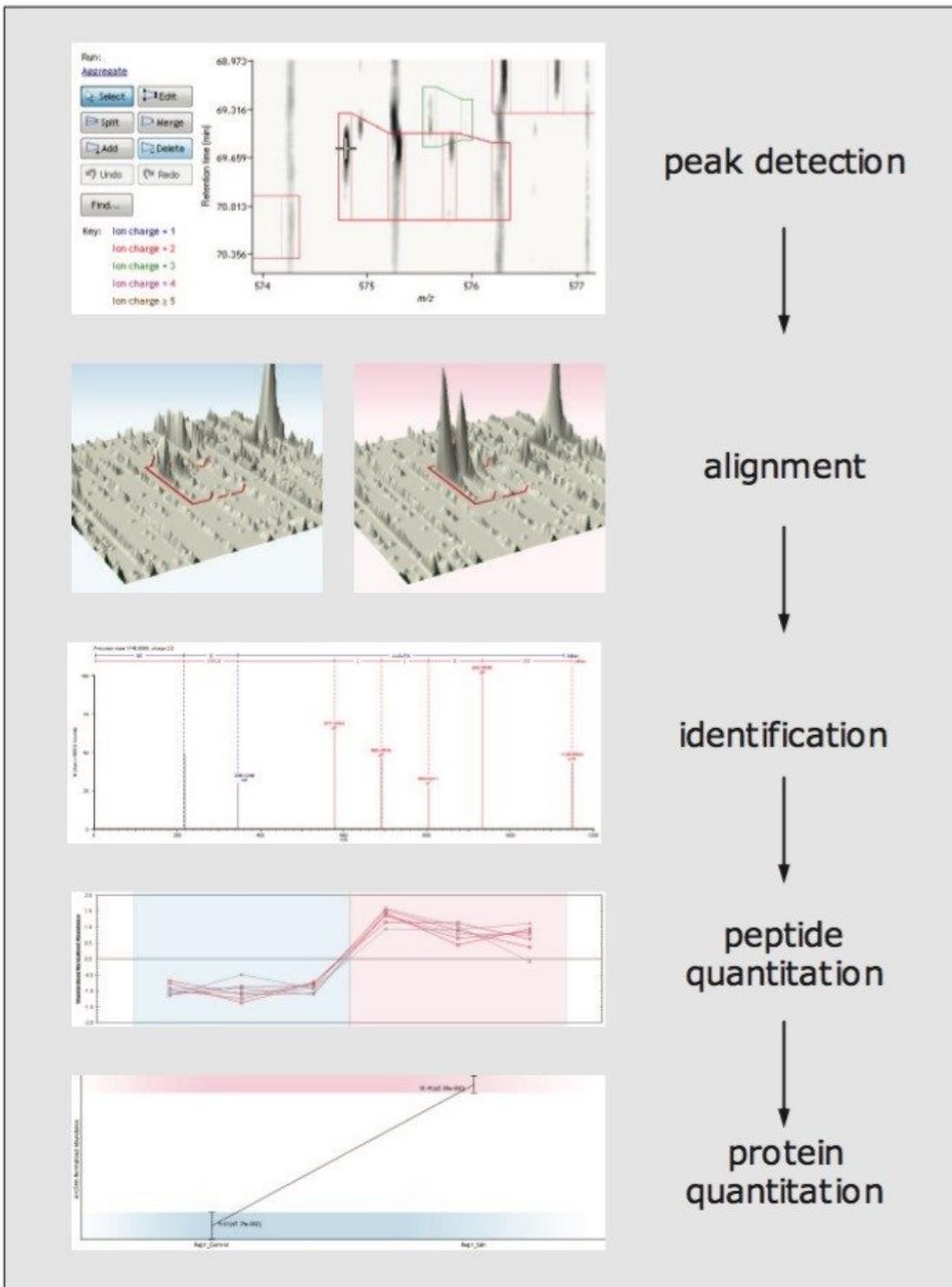


Figure 1. Workflow for Progenesis Q1 for proteomics.

Most of the data were identified in all samples using tolerances for matching peaks of $m/z \pm 5$ ppm, $t_r \pm 0.5$ min, and $t_d \pm 5\%$ units, as shown in the left-hand pane of Figure 2. The top 95% raw-abundance percentile of the complete data set was considered. To improve detection across injections and samples, alignment and co-detection of peaks were conducted and an aggregate constructed. The detection boundaries of the latter are passed back to individual samples, affording a complete data matrix that improves downstream analysis via multivariate statistics.

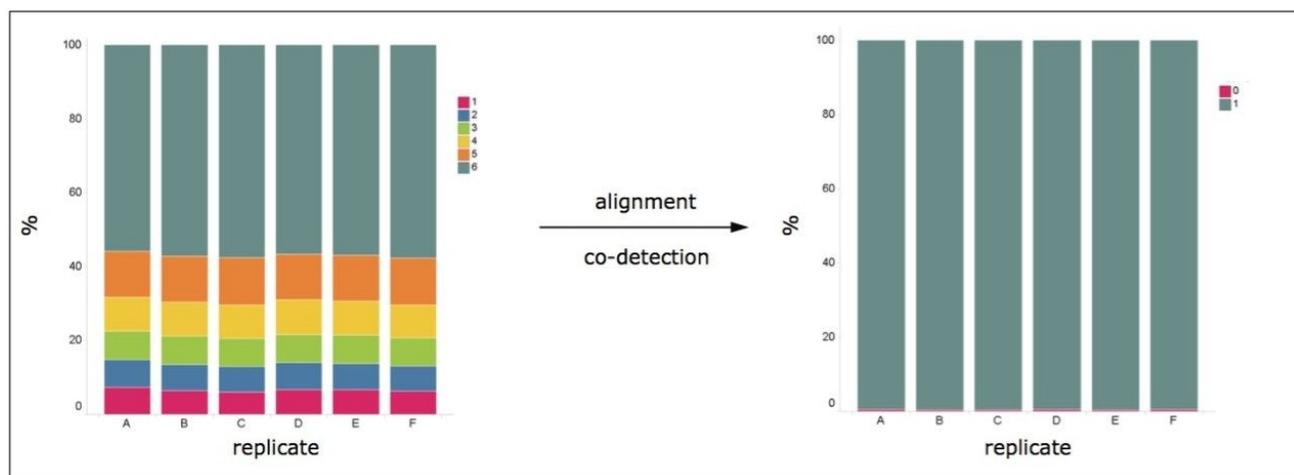


Figure 2. Percentage of features in each sample detected in all other samples (left) and percentage of features matched in the aggregate (right) for six, technical, LC-IM-DIA-MS replicates of *E. coli*. The light-grey color represents the features identified in all replicates.

Applying this principle and the same match criteria as that used for the previously described one-to-one replicate comparisons, the vast majority of the detected features in the individual runs could be identified in the aggregate (right-hand pane of Figure 2). Comparing individual runs, approximately 55% of the features could be detected in all runs, illustrated by the light-blue sections of the Venn intersection shown at the left-hand side of Figure 2. In contrast, nearly 100% of the features could be identified in the aggregate run, as illustrated by the Venn diagrams as illustrated at the right-hand side of Figure 2. An average increase of 98.3% in co-detected features was observed.

Three replicates of each *E. coli* sample, differentially spiked with BSA, ADH, enolase, and glycogen phosphorylase B, were also analyzed by mobility assisted, data-independent LC-MS. Figure 3 shows part of the

quantitative analysis of the data, including a results summary for the protein spikes, using ADH as the internal standard. All spikes were confidently quantified with expected ratios, as specified by the manufacturer.

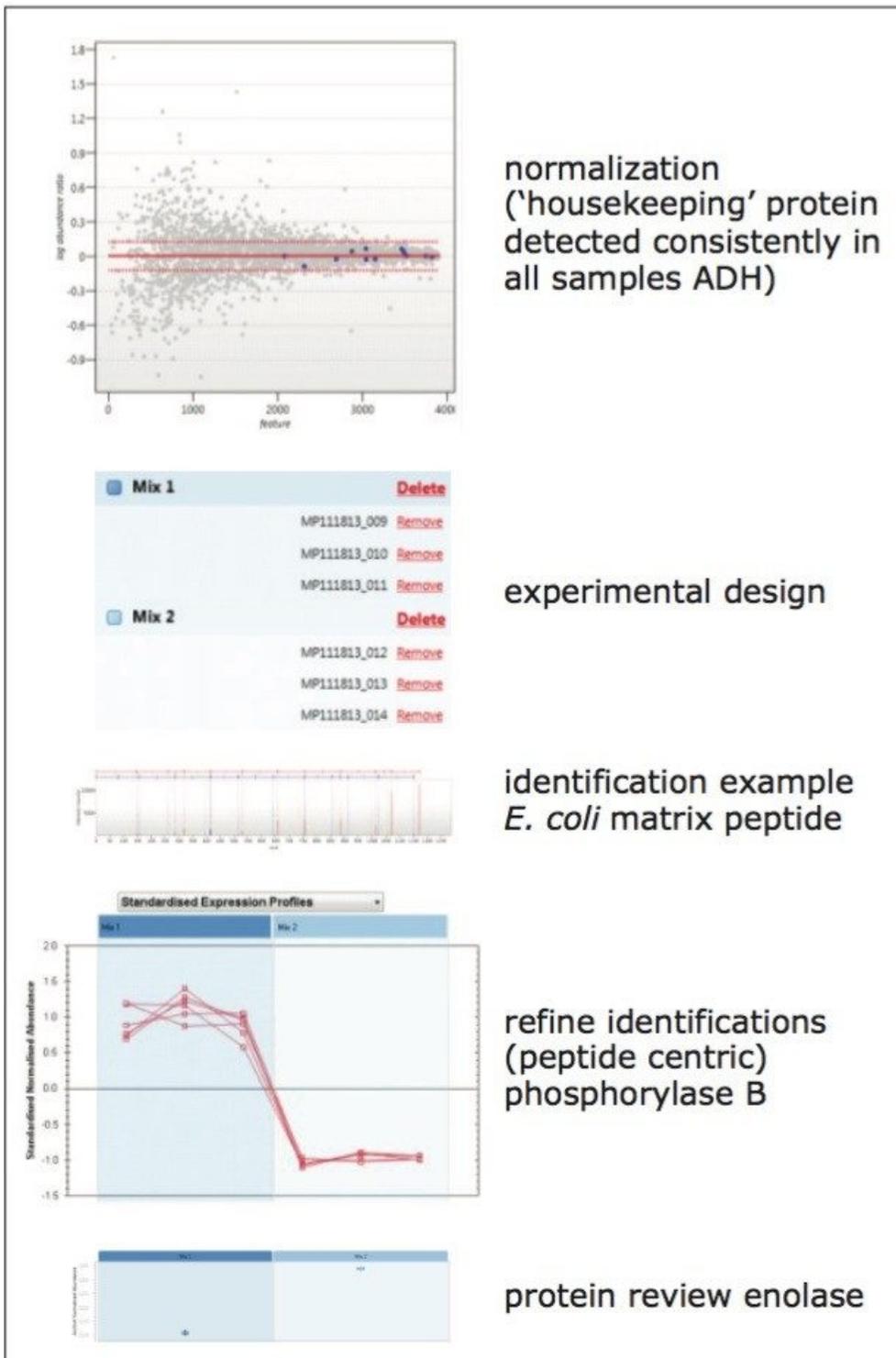


Figure 3. Workflow and quantitative results of a label-free LC-IM-DIA-MS experiment.

In quantitative workflows for which stable-isotope labeling is used, such as SILAC or dimethyl-labeling experiments, peptide pairs are expected to have similar retention and ion-mobility drift times. The results in Figure 4 illustrate the detection of a dimethyl-labeled peptide pair showing a mass spectrum detail (a), a section of the chromatographic separation with a partially resolved pair highlighted in red (b), and the ion-mobility separation (c) from a human cell-line sample. As expected for dimethyl-labeled peptides, the chromatographic apices are offset, but cross sections and drift times are very similar. Peptide and protein quantification was conducted with ProteoLabels.

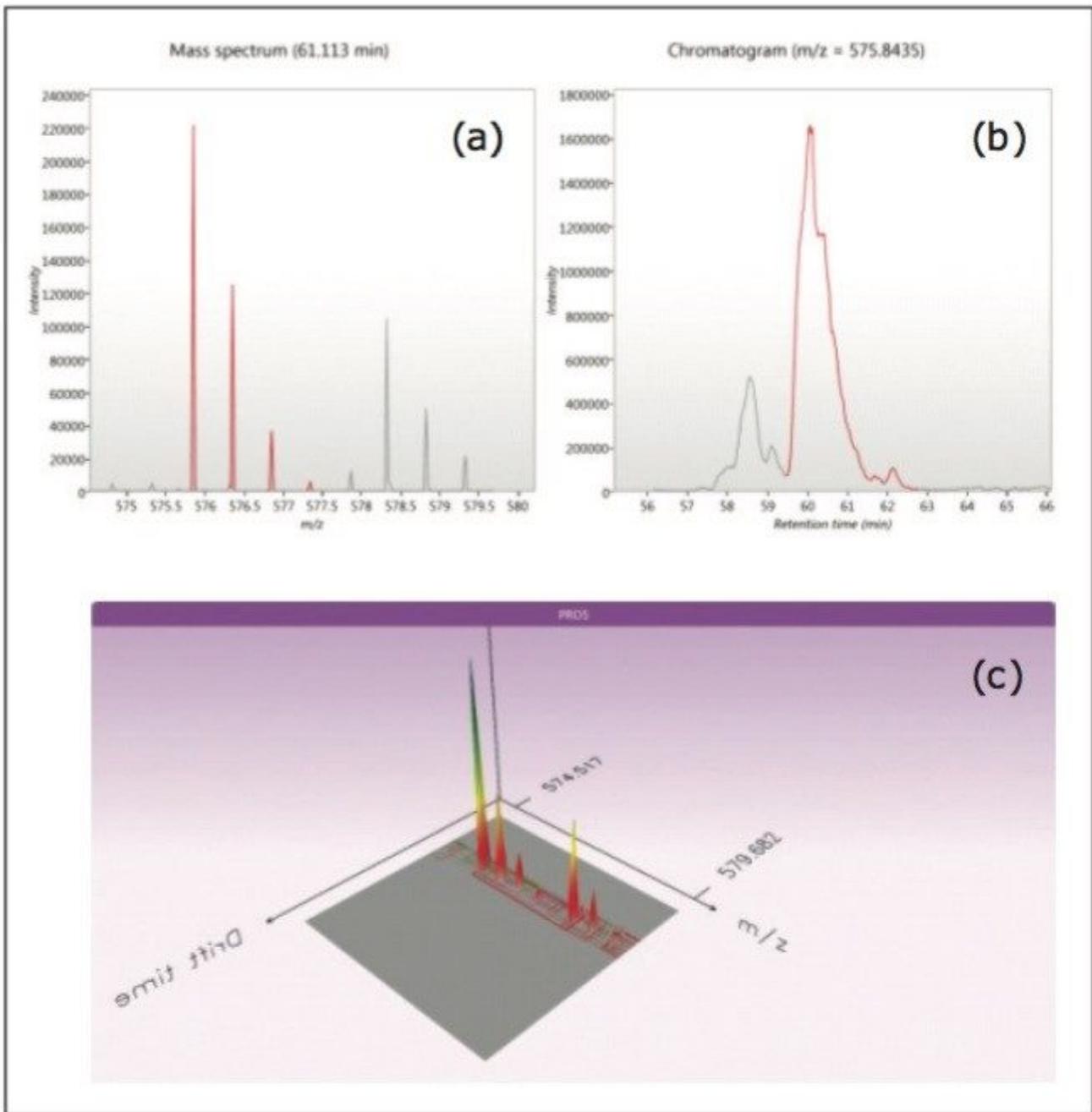


Figure 4. Detection (a,b) and IM separation (c) of a dimethylated peptide pair with a mass spectral detail, a section of the chromatographic separation illustrating a partially resolved pair, and the ion mobility separation of the pair.

Figure 5 shows three ProteoLabels visuals including dimethyl-pair identification mapped onto a m/z versus tr contour plot (a), three technical replicate overlaid ratio distributions (b), and an abundance ratio chart highlighting the precision of the replicate experiments. A quantitative, protein-centric graphical summary of the experiment is shown on the right, with Student's t-test p-value versus log2 fold change and the number of quantifiable peptides represented by size.

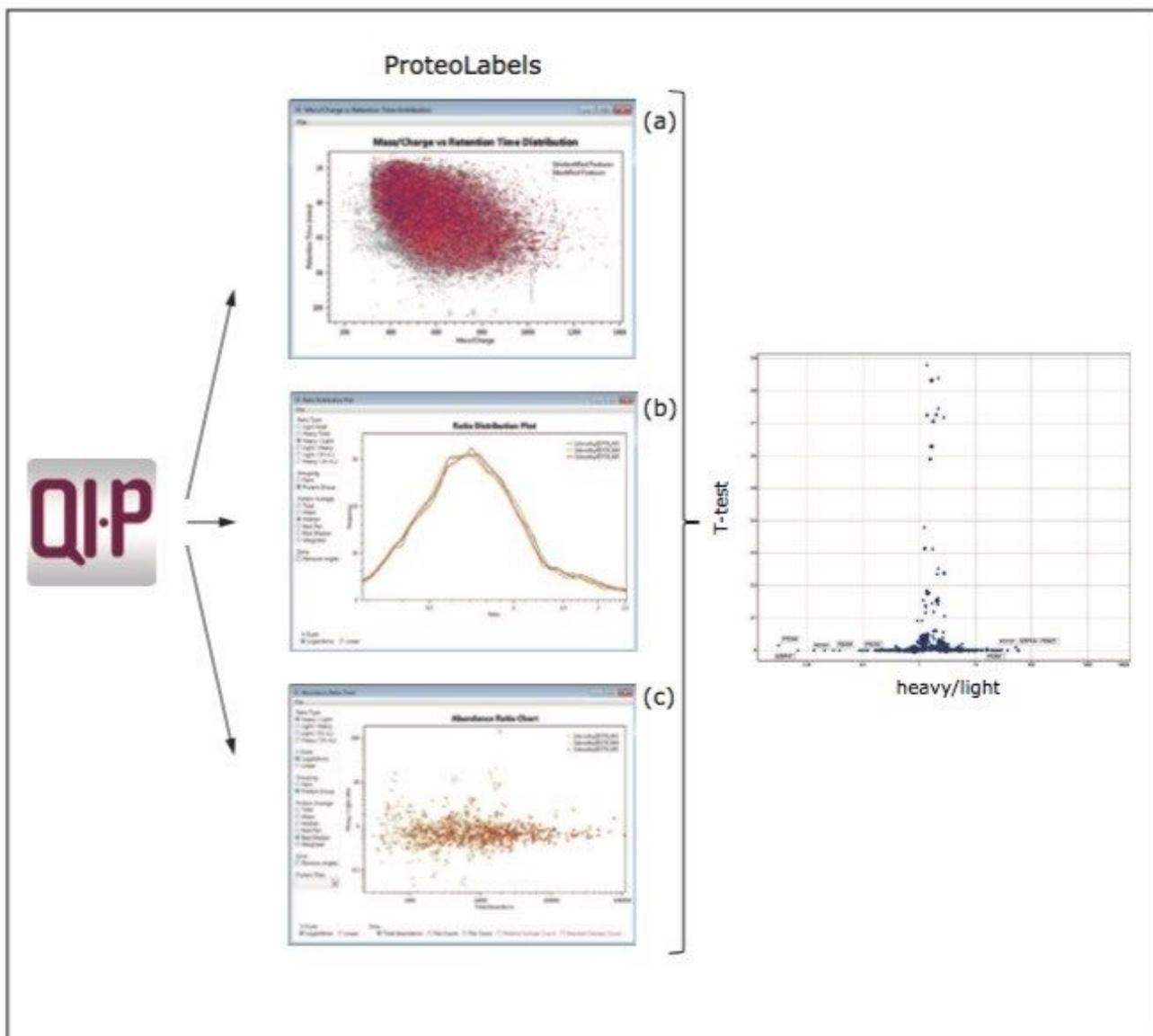


Figure 5. LC-IM-DIA-MS data analysis of dimethyl-labeled peptides and proteins following co-detection and peptide identification using Progenesis QI for proteomics (QI.P), Proteolabels pair analysis (middle figures a, b and c), and quantitation visulation (right).

Progenesis QI for proteomics also affords the label-free quantitation of DDA data. Figure 6 shows the detection and subsequent results for the labelfree quantification of one of the UPS1 standards (SYUG_HUMAN) that was differentially spiked in a tryptic digest of yeast, with on-column amounts of 25, 2.5, and 0.125 fmol, respectively, and analyzed by DDA. The isotopic clusters with peptide and protein distribution profiles are presented. Pane (a)

of Figure 6 illustrates the detection of one peptide precursor, with averages for the duplicates of the three measured samples. Pane (b) represents the intensity distribution profiles of all peptides identified to SYUG_HUMAN, with a protein-level summary, including variances, shown in pane (c).

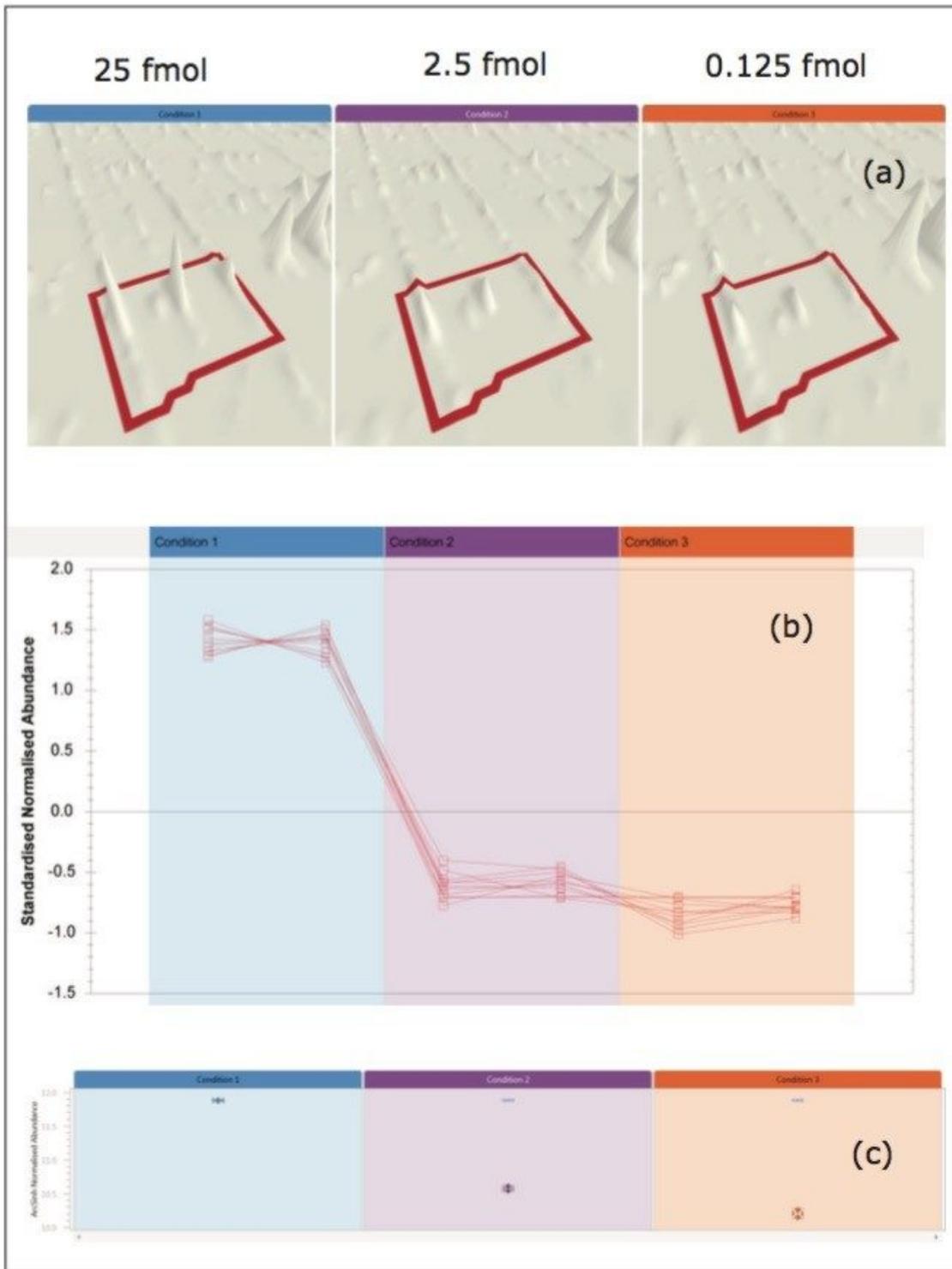


Figure 6. Quantitative label-free analysis DDA data of UPS1 standard Gamma-synuclein (SYUG_HUMAN),

showing feature detection (a), peptide quantitation (b), and protein quantitation (c) across three samples.

Conclusion

Progenesis QI for proteomics was successfully applied for a number of “bottom-up” proteomic applications, including the analysis of labeled and nonlabeled data acquired in DDA or DIA mode of acquisition. Moreover, consistent peak detection and the formation of an aggregate allowed for enhanced differential and statistical analysis. Lastly, the precision and accuracy of DIA and DDA quantitation were significantly improved using a co-detection based, label-free, quantitation approach.

Acknowledgements

Dimethyl-labeled HL60 human B cells were kindly donated by Bobby Florea, Bio organic Synthesis, Faculty of Science, Leiden Institute of Chemistry, the Netherlands.

ProteoLabels software was provided by Andrew Collins and Andy Jones from Institute of Integrative Biology, University of Liverpool, United Kingdom.

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720005239, December 2014

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