

A Facile Database Search Engine for Metabolite Identification and Biomarker Discovery in Metabolomics

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Abstract

In this study, we show the Progenesis QI workflow for metabolite identification using, as an example, a study on the effect of different bottling conditions on the nutritional composition of Italian wines.

Progenesis QI effectively streamlines and simplifies complicated metabolomics workflows and makes metabolite identification faster, easier, and more robust. User-definable search parameters dramatically decrease the number of false positive and false negative results in the identification workflow, improving the confidence of identification.

Benefits

Progenesis QI Informatics simplifies the process of metabolite identification and biomarker discovery. Potential biomarkers can be searched in both publicly available and in-house databases for accurate mass, retention time, collision cross section, and fragmentation information. Such an approach is fast and it increases the confidence of metabolite identification in metabolomics experiments.

Introduction

Metabolomics experiments offer a promising strategy for biomarker discovery. In a metabolomics workflow, however, the major bottleneck still remains metabolite identification. Currently, there are four levels of annotation for metabolite identification: 1) Confidently identified compound (two orthogonal properties based in authentic chemical standard analysis under the same condition); 2) Putative identified compounds (one or two orthogonal properties based in public database); 3) Putative identified compound class; and 4) Unknown compound.¹ A typical database search that relies only on one property (i.e., accurate mass) usually leads to an extensive number of false positive and negative identifications. To increase the confidence of identification, a search engine should be able to use of in-house databases containing orthogonal molecular descriptors for each metabolite.²

Progenesis QI Informatics is a novel software platform that is able to perform alignment, peak-picking, and mining of metabolomics data to quantify and then identify significant molecular alterations between groups of samples. The software uses a search engine (MetaScope) for metabolite identification, with user-definable search parameters to probe both in-house and publicly available databases. With an easy-to-use interface, the user can combine information for metabolite identification, including accurate mass, retention time, collision cross section, and theoretical and/or experimental fragment ions. These physiochemical properties can increase the confidence of metabolite identification while concurrently decreasing the number of false positives.

In this study, we show the Progenesis QI workflow for metabolite identification using, as an example, a study on the effect of different bottling conditions on the nutritional composition of Italian wines.

Experimental

UPLC conditions

System:	ACQUITY
	UPLC

Column: ACQUITY
UPLC BEH
HSS T3 1.8 μ
m 2.1 x 150
mm (p/n
186004120)

Pre column: ACQUITY
UPLC BEH
HSS T3
VanGuard
1.8 μ m, 2.1 x
5 mm
(p/n186003976)

Mobile
phase A: Water +
0.1% formic
acid

Mobile
phase B: Methanol +
0.1% formic
acid

Flow rate: 0.28 mL/min

Column
temp.: 40 °C

Injection
volume: 10.0 μ L

Gradient:

Min	A%	B%	Curve
Initial	100.0	0.0	Initial
1.0	100.0	0.0	6.0
3.0	90.0	10.0	6.0
18.0	60.0	40.0	6.0
21.0	0.0	100.0	6.0
25.5	0.0	100.0	6.0
25.6	100.0	0.0	6.0
28.0	100.0	0.0	6.0

MS conditions

MS system:	SYNAPT HDMS
Mode of operation:	Tof MS ^E
Ionization:	ESI +/-
Capillary voltage:	2.5 kV (+) and 2.5 kV (-)
Cone voltage:	25 V
Transfer CE:	Ramp 15 to 45 V

Source temp.:	150.0 °C
Desolvation gas flow:	1000 L/h
Desolvation temp.:	500.0 °C
Cone gas:	50 L/h
MS gas:	Nitrogen
Acquisition range:	50 to 1200

Data processing and mining:

Progenesis QI Informatics

Sample collection and preparation

Mezzacorona winery (Trentino, Italy) provided the wines, which were bottled in typical 750-mL wine bottles with the filling industrial machine of the winery. The sample set included two types of wines bottled with nitrogen addition (N₂) and without nitrogen addition (O₂). Under nitrogen atmosphere, every wine was uncorked, 2 mL were transferred into a 5-mL amber vial, 2 mL Milli-Q water was added, and finally each sample was filtrated with 0.2-µm PTFE filters into a 2-mL Waters LCMS Certified Amber Glass Vial prior to LC-MS analysis.³

Results and Discussion

Wine is one of the most complex foods as far as its metabolomic profile is concerned, since grapes, yeasts, bacteria, fungi, exogenous antioxidants, fining agents, and other oenological materials, packaging, and aging are involved in its preparation. This great number of different primary and secondary metabolites, most of which are unknowns, highly affects wine quality and the important role it plays in human diet, health, and enjoyment. Different bottling and storage conditions may affect the molecular composition of wines, and thus value and

quality.

In this study, we used Progenesis Q1 to identify the metabolites that were altered between wines bottled under two different levels of oxygen: high level (O₂) versus low level (N₂). Data were acquired in LC-MS^E mode (Figure 1A) and pre-processed using retention time alignment and peak picking (Figure 1B). A composite ion map was built, which contained more than 3,000 compounds after isotopic and adduct deconvolution (Figure 1B).

Metabolites of interest were filtered according to the ANOVA P value <0.01 and fold change >2, which decreased the number of metabolites of interest (markers) to less than 200 (Figure 2A). This data reduction strategy allowed us to focus on the metabolites that clearly discriminate the two groups of samples as shown by principal component analysis (Figure 2B).

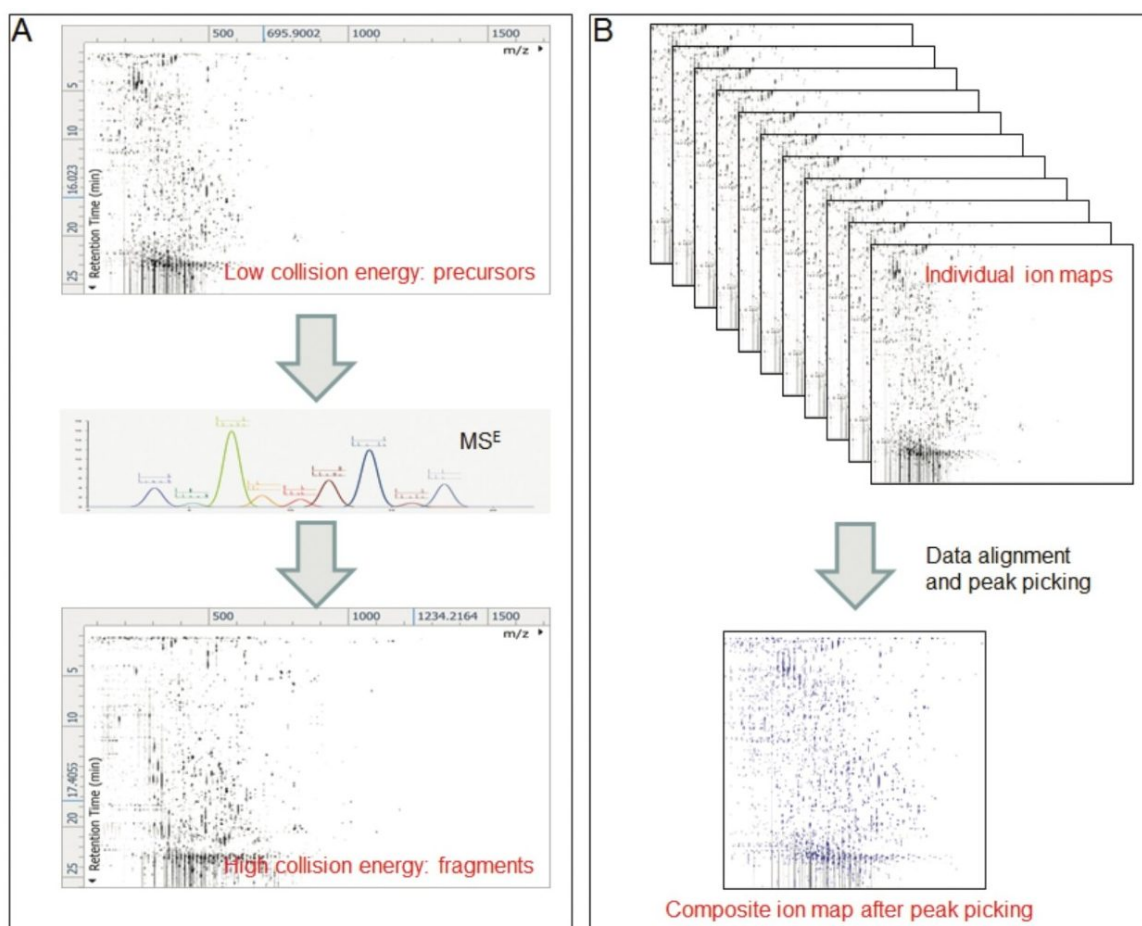


Figure 1. A: Samples were acquired using data independent analysis (MS^E), which provided information for both the intact precursor ions (at low collision energy, upper panel) and the fragment ions (high collision energy, bottom panel). B: From the aligned runs, Progenesis Q1 produces an aggregate run that is representative of the compounds in all samples, and uses this aggregate run for peak picking. The peak picking from this aggregate is then propagated to all runs, so that the same ions are detected in every run.

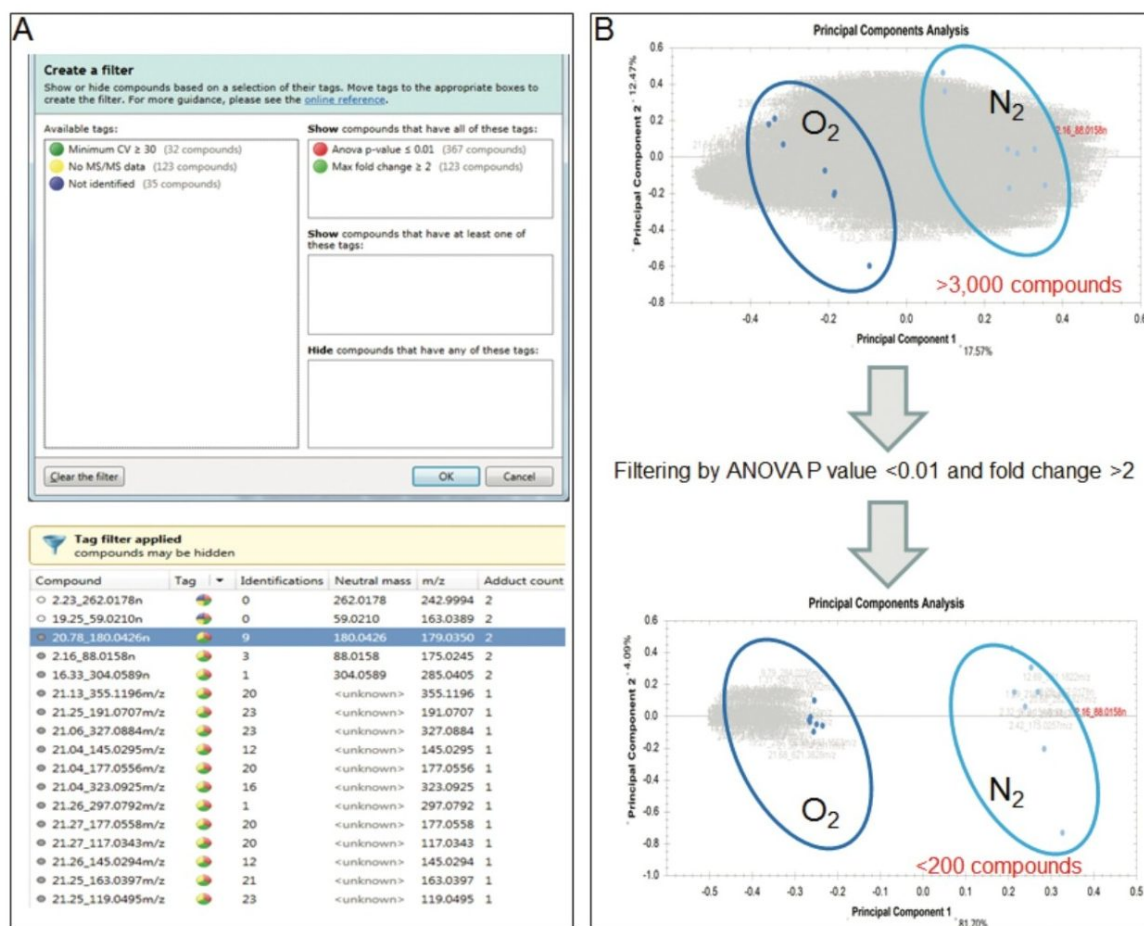


Figure 2. A: Progenesis Q1 allows tagging data according to various criteria, including ANOVA P values and fold changes. B: Principal component analysis (PCA) containing the entire dataset showed that the wines samples clustered according to the different amount of oxygen in which they were stored, suggesting that the two groups of wine contained a diverse set of metabolites (upper panel). After data reduction, PCA showed the discriminatory power of the selected compounds by filtering only those compounds that had ANOVA p values <0.01 and fold changes >2 (bottom panel).

Initial identification of metabolites was performed using the Human Metabolome Database (HMDB), leading to multiple ambiguous identifications for each compound of interest (Figure 3A and 3B). To decrease the number of false positives, we used in-house metabolite databases, which contain accurate mass, retention time, and fragment information.² (Figure 3A-C). We customized the search engine parameters for these orthogonal

measures (Figure 3A), allowing a more balanced set of tolerance criteria, which significantly decreased the number of false positives and false negatives (Figure 3B). Experimental fragments were matched against those derived from theoretical fragmentation to further increase the confidence in metabolite identification (Figure 3C). The entire metabolomics workflow for data processing, mining, and identification was completed in just a few hours.

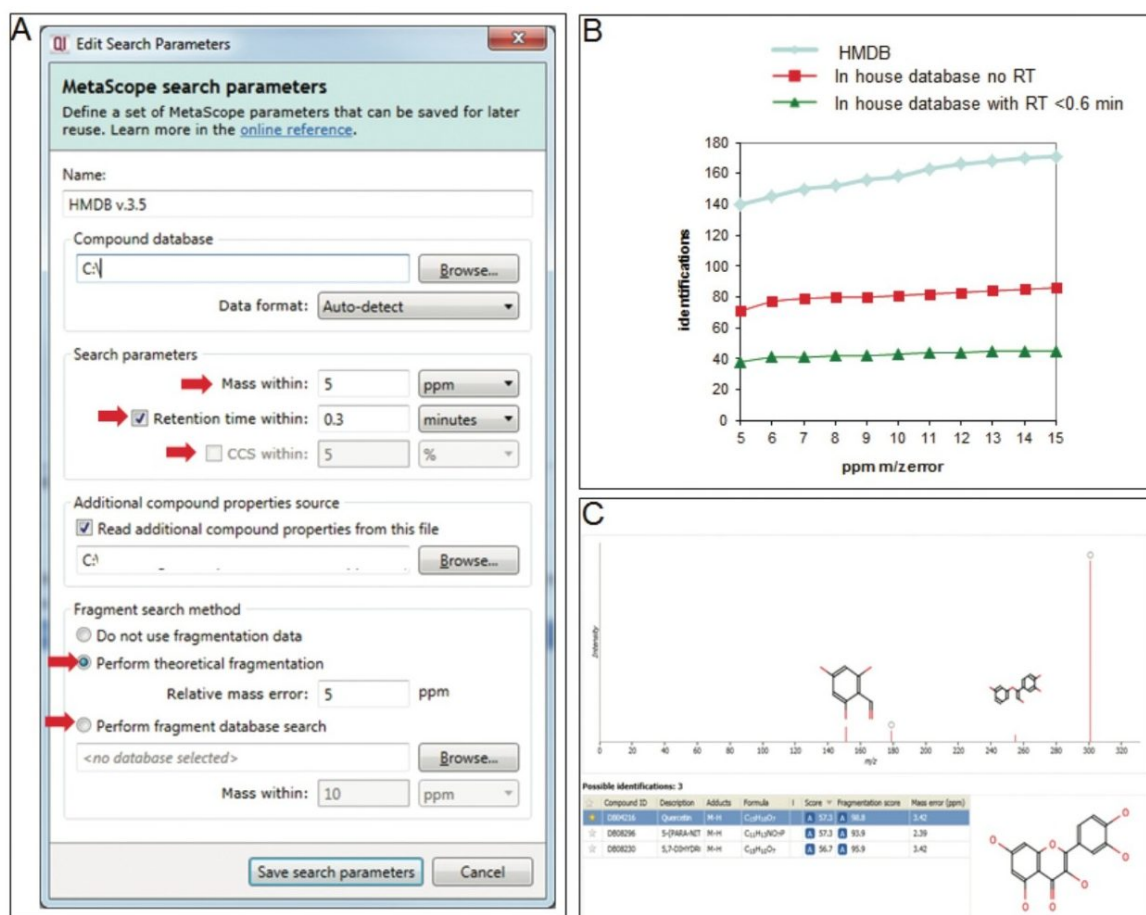


Figure 3. A: The Progenesis Q1 search engine allows users to query both publicly available databases (e.g., HMDB) and in-house databases, customizing the search parameters for the metabolite identification according to multiple orthogonal measures: mass accuracy, retention time, collision cross section and fragmentation matching. B: Metabolite identification using in-house database allows to filter the results by mass accuracy and retention time tolerance reducing significantly the number of false positives. C: Representative identification of the metabolite Quercetin using mass accuracy, retention time, isotopic distribution and four MS^E fragments, which were matched against theoretically-generated fragments.

Conclusion

Progenesis QI effectively streamlines and simplifies complicated metabolomics workflows and makes metabolite identification faster, easier, and more robust. User-definable search parameters dramatically decrease the number of false positive and false negative results in the identification workflow, improving the confidence of identification.

References

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