

# Fast and Sensitive UPLC-MS/MS Determination of Diquat and Paraquat in Drinking Water

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### Abstract

This application note describes an optimized method for determination of paraquat and diquat in drinking water using an Oasis WCX cartridge for SPE prior to fast and sensitive UPLC-MS/MS analysis.

#### Benefits

- Oasis WCX SPE cartridges incorporate a mixed-mode weak cation-exchange sorbent that is ideal for retention of quaternary ammonium compounds such as paraquat and diquat.
- UPLC-MS/MS with the ACQUITY BEH HILIC column provides a rapid and sensitive analytical method without the need for ion-pairing reagents that can be detrimental to good mass-spectral response.
- The LOQ for this method is 40 ng/L (parts per trillion).

#### Introduction

Paraquat and diquat are quaternary ammonium herbicides used in great quantities worldwide for defoliation and weed control (see Figure 1). In the United States alone, paraquat use was 4 million pounds and diquat use was 20,000 pounds in 2002<sup>1</sup>. Because these compounds are potentially toxic to wildlife and to humans, groundwater and surface water samples are monitored to assure that residue levels meet safety standards. These limits range from 100 ng/L in Europe to 20 µg/L in the United States. This application note describes an optimized method for determination of paraquat and diquat in drinking water using an Oasis WCX cartridge for SPE prior to fast and sensitive UPLC-MS/MS analysis. Oasis WCX is a mixed-mode weak cation exchange sorbent that is effective for isolation of paraquat and diquat from aqueous samples. A prior publication discussed HPLC-MS determination of quats after SPE with Oasis WCX<sup>2</sup>. The new UPLC method requires half the analysis time compared with the HPLC method. Also, the SPE has been improved; the earlier protocol utilized trifluoroacetic acid, a toxic perfluorinated compound and persistant organic pollutant. In the new SPE protocol, formic acid is used as an acidic modifier for the SPE eluent.



Figure 1. Structures for Diquat and Paraquat.

## Experimental

## LC conditions

LC System:	ACQUITY UPLC H-Class system
Column:	ACQUITY BEH HILIC
Mobile Phase (Isocratic):	40:60 A/B
Mobile Phase A:	150 mM ammonium formate buffer (pH 3.7)
Mobile Phase B:	Acetonitrile
Injection Volume:	10 µL
Column Temperature:	30 °C
Weak Needle Wash:	50:50 methanol/water (600 µL)
Strong Needle Wash:	50:50 methanol/water (600 µL)
Seal Wash:	10:90 acetonitrile/water
Flow Rate:	0.50 mL/min
MS conditions	
MS System:	TQD
Ionization Mode:	Positive Electrospray
Source Temperature:	150 °C

Desolvation Temperature:	350 °C
Desolvation Gas Flow:	800 L/Hr
Cone Gas Flow:	30 L/Hr
Collision Gas Flow:	0.20 mL/Min
Data Management:	MassLynx v4.1

#### Experimental

Three types of water samples were chosen to demonstrate this new method. Chlorinated tap water was obtained from a local municipal source, groundwater was obtained from a commercial bottled source, and untreated surface water was obtained from a local river. All samples were adjusted with phosphate buffer to pH 7 prior to SPE. Municipal drinking water samples were dechlorinated with sodium thiosulfate prior to SPE. The Oasis WCX cartridge was then used for effective enrichment and cleanup prior to UPLC/MS(MS) analysis.

#### MRM transitions and LC-MS parameters

Compound	MRM	Cone (V)	CID (eV)
Diquat	183.1 > 157.1	50	25
	183.1 > 130.1	50	30
d4-Diquat	186.1 > 158.1	43	30
Paraquat	185.1 > 170.1	38	22
	171.1 > 77.0	45	40
d8-Paraquat	83.2 > 178.1	35	25

#### Reagents

#### pH adjustment buffer concentrate (400 mM phosphate buffer pH 7)

Accurately weigh 20.9 g of potassium phosphate dibasic and 10.9 g of potassium phosphate monobasic into a 500 mL volumetric flask. Add sufficient reagent water (Milli-Q or equivalent) to completely dissolve and then dilute to the mark reagent water. Check pH—if necessary, adjust to 7.0 using very dilute KOH or  $H_3PO_4$  aqueous solution.

#### SPE Eluent

Mix 200 mL acetonitrile, 200 mL isopropanol, and 100 mL formic acid (caution—use only with appropriate personal protective clothing and eyewear).

#### pH 7 phosphate buffer (25 mM) for SPE

Take 25 mL of the 400 mM pH 7 buffer and dilute to 400 mL with reagent water.

#### pH 8 formate buffer for SPE

Accurately weigh 1.6 g of ammonium formate into a 1 L volumetric flask. Add sufficient reagent water to completely dissolve and dilute to approximately 900 mL. Add 1.4 mL of 30 % ammonium hydroxide and mix well. Adjust to exactly pH 8 with dilute formic acid or ammonium hydroxide as needed. Dilute to mark with reagent water.

#### Sample preparation

Note: Polypropylene containers should be used for sample collection and for all sample preparation steps. Polypropylene autosampler vials (part number 186002642) are recommended for UPLC analysis.

#### 1. Sample pretreatment

Transfer a 10 mL sample to an appropriate polypropylene container (15 mL centrifuge tubes were used for this study). For chlorinated samples add 10 mg of sodium thiosulfate and mix well. For all samples, adjust pH by addition of 25 µL of 400 mM pH 7 phosphate buffer.

#### 2. SPE enrichment and cleanup:

Perform SPE enrichment and cleanup using Oasis WCX cartridges (see SPE details in Figure 2). To allow convenient loading of the 10 mL sample, attach a 30 cc polypropylene reservoir (part number WAT011390) to each cartridge.

# OASIS WCX, 3 cc 60mg, 30 µm

# (part number 186002495)

# CONDITION/EQUILIBRATE

3 mL methanol 3 mL 25 mM pH 7 phosphate buffer



# 500 µL 40:40:20

acetonitrile/IPA/formic acid

Figure 2. Oasis

WCX Cartridge Protocol for Diquat/Paraquat Analysis.

**Results and Discussion** 

Figure 3 shows typical LC-MS chromatograms obtained from analysis of a matrix-matched standard of diquat and paraquat at 40 ng/L prepared in dechlorinated municipal water. Performance was similar for the other water types. Table 2 shows the recovery data obtained from replicate analysis of water samples spiked at 40 ng/L. Typical matrix-matched calibration curves were linear in the range from 40–2000 ng/L for all water types. Representative calibration curves are presented in Figure 4.



Figure 3. Typical LC-MS/MS Chromatograms.



#### Diquat/paraquat recovery data (n=7) for three types of water spiked at 40 ng/L

Sample Type	Recovery (%RSD) Diquat	Recovery (% RSD) Paraquat
Groundwater	82 (8)	89 (16)
Tapwater	84 (5)	88 (8)
River water	83 (4)	89 (19)

#### Diquat

Correlation Coefficient: r = 0.996129, r<sup>2</sup> = 0.992272 Calibration Curve: 0.000420002 \* x + -0.00197468 Response Type: Internal Std ( Ref 3 ), Area\* (*IS* Conc./*IS* Area) Curve Type: Linear, Origin: Include, Weighting: 1/x, Axis trans: None



Figure 4. Typical LC-MS/MS Calibration Curves.

#### Conclusion

- Oasis WCX cartridges are effective SPE devices for the isolation of paraquat and diquat from aqueous samples.
- The paraquat/diquat SPE method is effective for enrichment and cleanup from various types of drinking water.
- · The UPLC-MS/MS method is fast, sensitive and requires no ion-pairing reagents.
- · Method recovery was better than 80% and ion-suppression was less than 30% for all types of water tested.

### References

1. Pesticide National Synthesis Project, 2002 Pesticide Use Maps, USGS

#### Paraquat

Correlation Coefficient: r = 0.995100, r<sup>2</sup> = 0.990223 Calibration Curve: 0.000575997 \* x + -0.000150743 Response Type: Internal Std ( Ref 4), Area\* (*IS* Conc./*IS* Area) Curve Type: Linear, Origin: Exclude, Weighting: 1/x, Axis trans: None



 M.S. Young and K. M. Jenkins, Oasis WCX: A Novel Mixed-Mode SPE Sorbent for LC-MS Determination of Paraquat and OtherQuaternary Ammonium Compounds, LC-GC, The Application Notebook, September 2004 (p 2-3)

## Featured Products

- ACQUITY UPLC H-Class Plus System <https://www.waters.com/10138533>
- MassLynx Mass Spectrometry Software <a href="https://www.waters.com/513164">https://www.waters.com/513164</a>>
- Xevo TQD Triple Quadrupole Mass Spectrometry <a href="https://www.waters.com/134608730">https://www.waters.com/134608730</a>

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