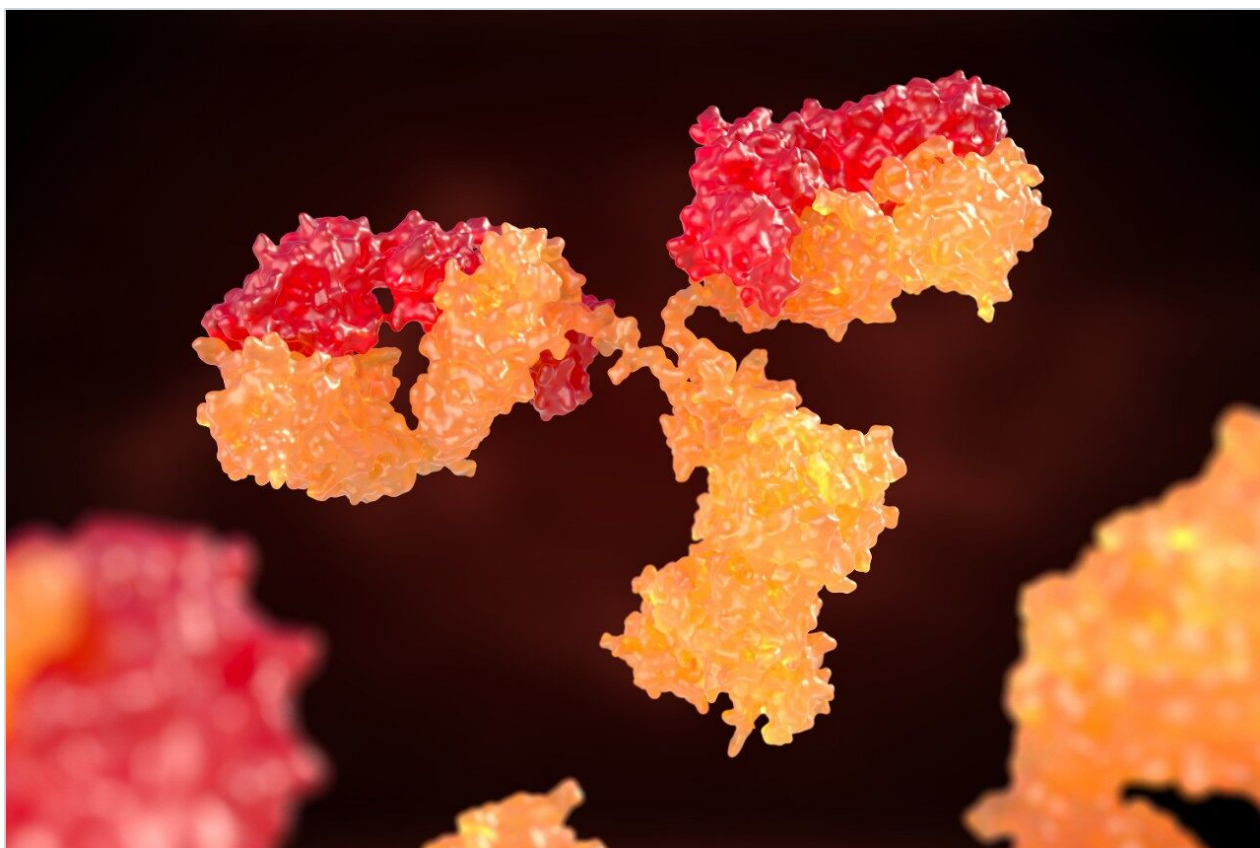


Resolving Chimeric Spectra Utilizing a Data Independent Mobility Acquisition Strategy

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This is an Application Brief and does not contain a detailed Experimental section.

Abstract

The specificity of a mobility assisted data independent acquisition (HDMS^E) in terms of resolving chimeric fragment ion spectra was explored through the analysis of a tryptic digested *Escherichia coli* lysate spiked with a labeled protein (bovine serum albumin), labeled with tandem mass tags (TMT).

Benefits

Analyze TMT labeled peptides with a high degree of specificity in a complex biological background using HDMS^E

Introduction

Protein identification and quantification are challenged by the presence of contaminating peptides within the collision cell during the collision induced dissociation (CID) process, leading to mixed fragment ion spectra obtained via either data dependent or independent acquisition strategies, also referred to as chimericity. This often unnoticed loss in acquisition specificity can result into incorrect peptide and protein identifications – leading also to an increase in protein identification false discovery rate (FDR) – and subsequently incorrect quantification values. The number of chimeric events does not only increase proportionally with sample complexity, but also increases with improved instrument sensitivity and dynamic range. A proof-of-principle example will be used to demonstrate that the incorporation of ion mobility into the experiment can aid in unambiguously assigning product to precursor ions, greatly increasing specificity and reducing protein and peptide FDR, and false quantification rate (FQR).

Results and Discussion

A tryptic digest of an *Escherichia coli* (*E.coli*) lysate was spiked with TMT labeled bovine serum albumin (BSA) at various concentrations. The peptides were separated and analyzed using a nanoACQUITY UPLC System coupled to a SYNAPT G2 HDMS. The data were acquired in LC-HDMS^E mode – an unbiased mobility assisted TOF acquisition method – switching between low and elevated energy on alternate scans.

Subsequent correlation of precursor and product ions can then be achieved using both retention and drift time alignment. Searches were conducted with ProteinLynx GlobalSERVER using a species specific database to which sequence information of BSA and other contaminating species was appended. Searching the DB

with TMT as a fixed modification was expected to only identify BSA, whereas variable searches should identify both *E.coli* proteins and BSA. Moreover, acquisitions that were not supported by means of ion mobility separations are expected to produce fragment *E.coli* ion spectra comprising the TMT reporter ions of co-eluting BSA peptides.

Figure 1 illustrates the identification overview for an HDMS^E acquisition with the TMT modification set as a fixed modification for a condition combined amount of approximately 70 fmol labeled BSA added to 750 ng *E.coli*. In contrast the search results of the same data with TMT specified as a variable modification resulted in 434 *E.coli* protein identifications, confirming the expected specificity afforded by the acquisition method and applied search algorithm. The results shown in Figure 2 demonstrate that without mobility separation, *E.coli* fragment ion spectra can be incorrectly appended with TMT reporter ions originating from co-eluting labeled BSA peptides, illustrating the requirement for increased system peak capacity via orthogonal peptide separation. Moreover, the spectrum shown in the bottom pane of Figure 2 comprises other contaminating product ions that do not arise from the precursor of interest and could have lead to an incorrect assignment if the multiplexed nature of the data would not have been acknowledged by the deconvoluting and search algorithms.

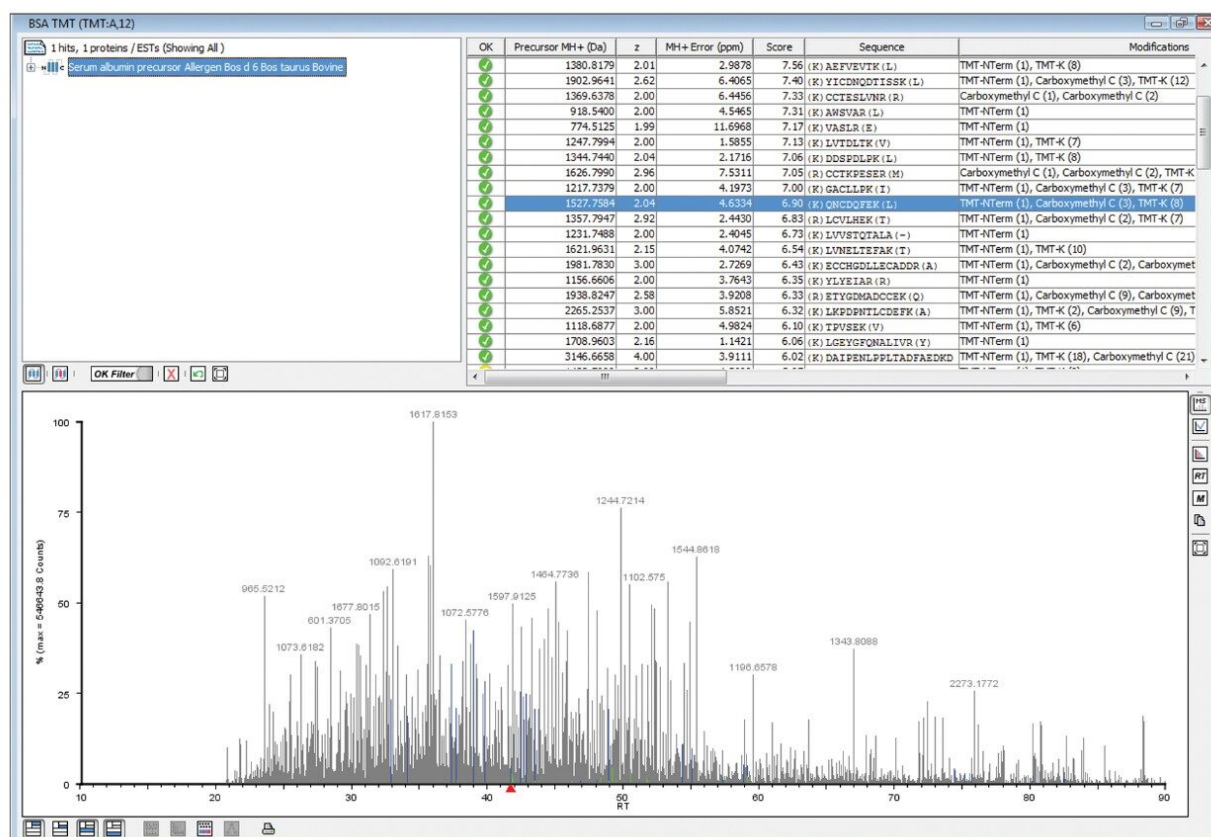


Figure 1. Protein identification overview of a condition combined amount of approximately 70 fmol labeled BSA added to 750 ng unlabeled E.coli with TMT set as a fixed modification. Color annotation peptides: blue = no or fixed modification and grey = non-identified peptide.

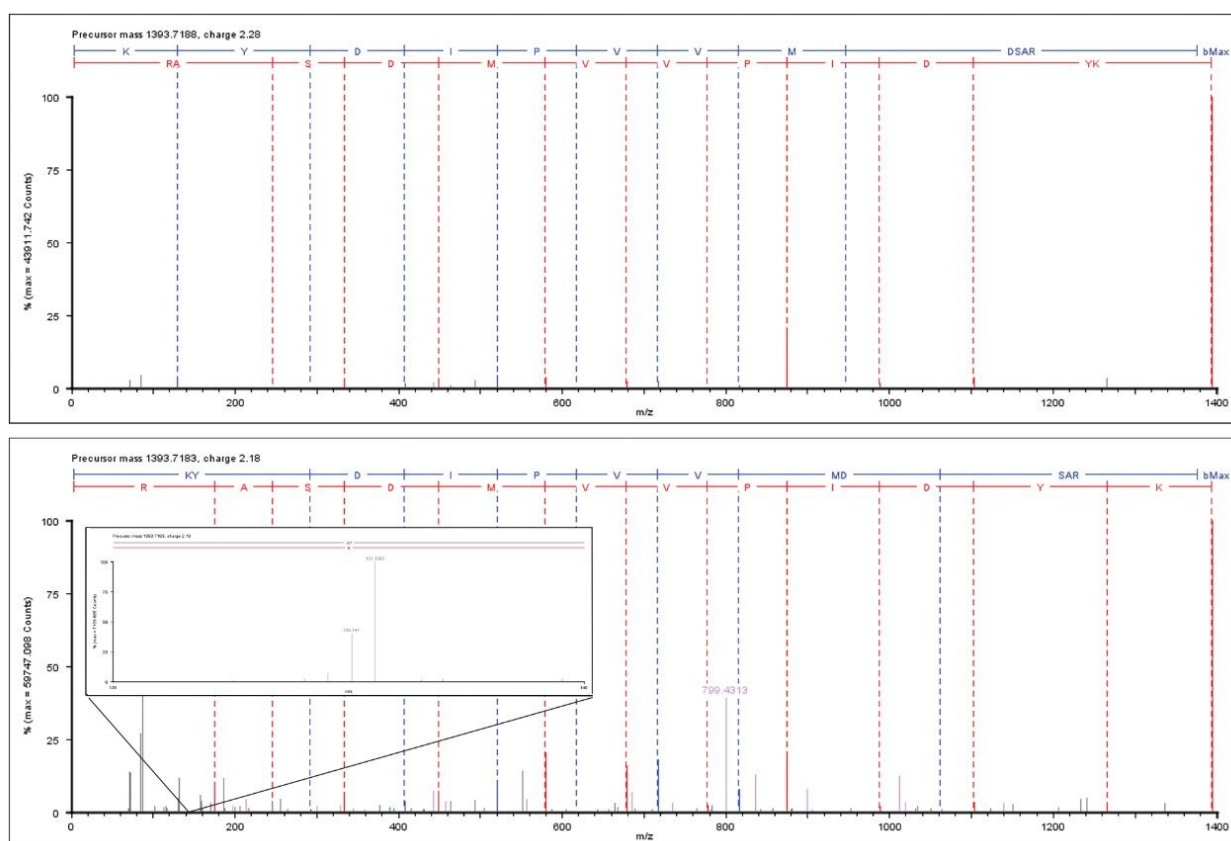


Figure 2. Data independent *Escherichia coli* fragment ion spectrum of KYDIPVVMDSAR – Tryptophanase with (top) en without (bottom) mobility separation and TMT set as a variable modification. Color annotation fragment ion: blue = b ion, red = yⁿ ion, magenta = fragment ion co-eluting peptide and grey = non-identified/assigned fragment ion.

Conclusion

The specificity of mobility assisted data independent acquisitions has been demonstrated for the analysis of TMT labeled peptides in a complex biological background. These results can be more specific than those obtained by more traditional data dependent acquisition (DDA) methods in terms of spectral fragment ion purity. The method additionally benefits from the orthogonal separation and correlation of precursor and product ions on the bases of their respective drift times and the high-resolution, accurate mass measurement of the product ions, all within a single LC-MS run.

Acknowledgement

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ACQUITY UPLC M-Class System <<https://www.waters.com/134776759>>

ProteinLynx Global SERVER (PLGS) <<https://www.waters.com/513821>>

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