

Rapid Analysis of Water-Soluble Vitamins in Infant Formula by Standard Addition

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Abstract

Vitamins are minor constituents that have to be introduced, via our food, in small quantities because they are not synthesized by the human body. The vitamin composition of infant formula is critical for correct infant development, particularly if the mother is unable to breast-feed and formula is the primary source of nutrition. Official analytical methods for the determination of water-soluble vitamins are based on procedures, mainly microbiological assays, which have been established for decades. Each vitamin is analyzed separately in order to apply extraction conditions, which permit the determination of its total content in a food. Vitamin analysis in food is generally a time-consuming process.

In this application note, we describe a rapid, five-minute UPLC-MS/MS method using positive ESI ionization for the simultaneous analysis of 12 water-soluble vitamin compounds in infant formula.

Benefits

This method allows for the simultaneous analysis of 12 water-soluble vitamin compounds:

- Replaces time-consuming microbiological assays of single compounds
 - Detects target compounds at low concentrations (particularly cyanocobalamin), in a very complex matrix, such as infant formula powder
 - Simultaneous acquisition of MRM and full scan data in a single analysis run
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Introduction

Vitamins are minor constituents that have to be introduced, via our food, in small quantities because they are not synthesized by the human body. The vitamin composition of infant formula is critical for correct infant development, particularly if the mother is unable to breast-feed and formula is the primary source of nutrition. Official analytical methods for the determination of water-soluble vitamins are based on procedures, mainly microbiological assays, which have been established for decades^{1,2}. Each vitamin is analyzed separately in order to apply extraction conditions, which permit the determination of its total content in a food. Vitamin analysis in food is generally a time-consuming process.

The development of a single method for their simultaneous determination of vitamins in fortified infant formula is difficult for several reasons:

- Diverse structures and chemical properties of the vitamin compounds
- Trace levels of vitamins present
- Matrix complexity
- Instability to light and heat
- Solubility issues
- Huge range of concentrations in infant formulas

In this application note, we describe a rapid, five-minute UPLC-MS/MS method using positive ESI ionization for the simultaneous analysis of 12 water-soluble vitamin compounds in infant formula.

Experimental

Throughout the sample preparation and analyses, all solutions were protected from exposure to light and stored at <5 °C.

Standard solutions of the vitamin compounds were prepared fresh daily.

LC conditions

LC System:	ACQUITY UPLC System
Column:	ACQUITY UPLC HSS T3, 1.8 μ m, 2.1 x 50 mm
Column temp:	40 °C
Sample temp:	4 °C
Flow rate:	0.6 mL/min
Mobile phase A:	10 mM Ammonium formate in water + 0.1% formic acid
Mobile phase B:	10 mM Ammonium formate in methanol + 0.1% formic acid
Total runtime:	5.0 min
Injection volume:	10 μ L, full loop

Gradient

Time (min)	%A	%B
0.0	99.0	1.0
2.0	99.0	1.0
3.0	45.0	55.0
3.1	1.0	99.0
4.0	99.0	1.0
5.0	99.0	1.0

MS conditions

MS System:	Xevo TQ MS
Ionization:	ESI positive
Capillary voltage:	1.0 kV
Source temp:	150 °C
Desolvation temp:	600 °C
Desolvation gas:	1200 L/hr
Acquisition:	Multiple Reaction Monitoring (MRM) with RADAR full scan Collision gas: Argon at 3.5×10^{-3} mbar

Standard-addition method

Since blank samples were not available, quantitative analysis of water soluble vitamins in infant formula powders was performed by the standard-addition method (except for nicotinamide)*. An analyte solution of known concentration (standard solution) was added to the sample so any matrix effects were accounted for in the calibration. The analyst did not know the amount of analyte in the sample initially, but tracked how much standard solution was added, and how the instrument response changed after adding the standard solution. Thus, by extrapolation of the calibration curve, the concentration of analyte in the sample could be determined. In practice, the volume of standard solution added is kept small to avoid dilution of the sample matrix.

* Using the current dilution factor (see experimental steps below) for the sample matrix without addition of standards, the instrument response of nicotinamide (B3) initially present in the infant formula is close to reaching the saturation of the detector. If standard-addition method is used for nicotinamide, detector saturation will be reached.

Sample preparation, extraction, and standard-addition

Prepare the infant formula to three times the concentration described on the packaging.

Place 10 mL of infant formula solution into a PP tube covered with foil; add 20 mL of 100% ethanol.

Shake vigorously for 2 min, and centrifuge for 15 min at 3500 RPM.

Filter supernatant using a 0.45 PVDF filter.

Transfer 20 μ L of the supernatant into an amber autosampler vial, add 10 μ L of known concentrations of standards containing 11 analytes, and top off the autosampler vial to 1 mL with water (the analyte in sample matrix was diluted 50 times; analyte in standard was diluted 100 times in final volume).

A separate standard curve of nicotinamide in solution was prepared.

Analyze by LC-MS/MS.

Acquisition and processing methods

Data were acquired using MassLynx Software, v.4.1, and processed using TargetLynx Application Manager.

IntelliStart Technology was used to automatically develop fully optimized MRM acquisition methods for the 12 vitamin compounds targeted in this analysis. IntelliStart requires only the entry of basic compound information, and automatically locates the precursor ion, optimizes cone voltage, locates product ions, and optimizes collision energy.

Two MRM transitions were optimized for each vitamin compound; the first transition for quantitation and the second transition for confirmation. The dwell times for the transitions were automatically optimized to give a minimum of 12 points across each chromatographic peak for reproducible quantitation. The MRM transitions, cone voltages, and collision energies for the analyzed compounds, along with expected retention times, are shown in Table 1.

Analyte		Parent (m/z)	Dau 1/ Dau 2 (m/z)	CV (V)	CE 1/ CE 2 (eV)	RT (min)
Ascorbic acid	C	177.0	141.0 95.0	16	8 12	0.37
Thiamine	B1	265.2	122.0 144.0	18	16 12	0.41
Nicotinic acid	B3	124.0	80.2 53.0	34	20 22	0.51
Pyridoxal	B6	168.0	150.0 94.0	14	14 24	0.64
Pyridoxine	B6	170.0	152.0 134.0	20	12 20	0.86
Nicotinamide	B3	123.0	80.0 106.0	32	18 12	0.93
Pantothenic acid	B5	220.1	90.0 202.1	20	14 12	2.73
Cyanocobalamin	B12	678.6	147.1 359.2	30	36 24	2.98
Folic acid	B9	442.2	295.1 176.0	18	16 36	2.99
Riboflavin-5'-phosphate	B2	457.2	439.2 359.2	30	16 20	3.04
Biotin	B7	245.1	227.0 97.0	20	14 30	3.10
Riboflavin	B2	377.2	243.1 172.1	36	24 42	3.15

Table 1. LC-MS/MS parameters for the identification of water-soluble vitamin compounds.

In addition to MRM data, full scan data were acquired using the RADAR mode of the Xevo TQ MS. RADAR is an information-rich acquisition approach that enables real time acquisition of spectral information on background components in the sample matrix, while simultaneously collecting MRM data. The use of RADAR does not compromise the quality of the MRM data.

Results and Discussion

12 water-soluble vitamin compounds were successfully analyzed using the ACQUITY UPLC System, coupled

with Xevo TQ MS under ESI positive ionization. The use of ACQUITY UPLC enabled rapid separation of all analytes in <5 min, including 1 min for equilibration, as shown in Figure 1.

In the same analysis, full scan spectra to assess the background components in the infant formula matrix were also monitored using the RADAR mode of the Xevo TQ MS. RADAR utilizes the fast acquisition rates of the Xevo TQ MS, allowing full scan MS data to be acquired, while still collecting a sufficient number of points across the analyte peak, in MRM mode, for accurate quantification and confirmation.

With RADAR, the analyst can observe untargeted contaminants in the sample matrix, and get an idea of the level and type of compounds causing possible matrix effects. This provides insight in the development of matrix reduction strategies during LC-MS/MS methods development. For example, the separation method could be modified to move the peaks of interest away from where areas of potential matrix effects or ion suppression are likely to be present. An illustration of a MS spectrum extracted from a section of the full scan data of the infant formula sample matrix is shown in the insert of Figure 1.

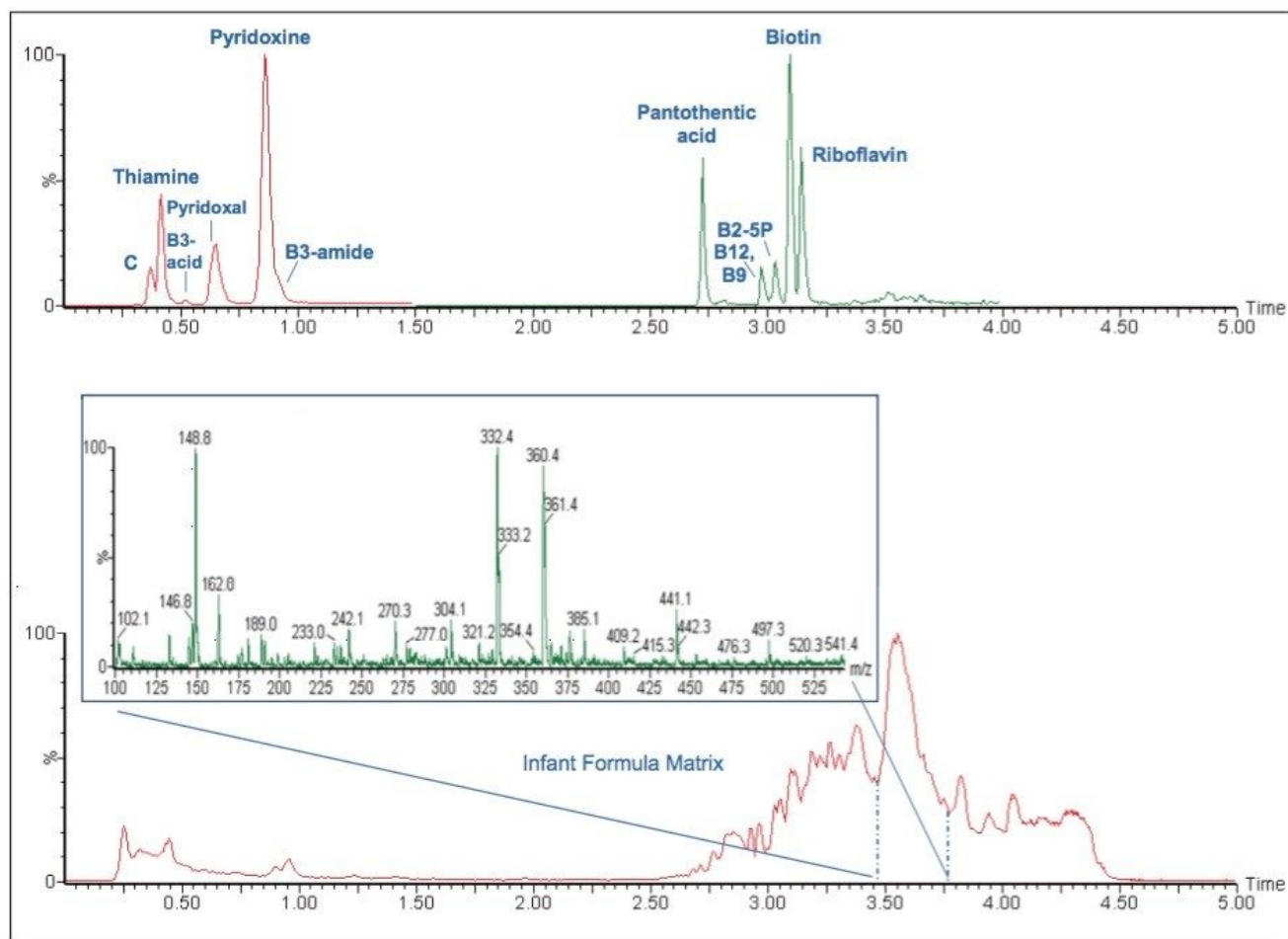


Figure 1. RADAR technology allows for simultaneous acquisition of MRM and full scan data in a single analysis run. Spectrum identifying untargeted contaminants in sample matrix are shown in the insert of the full scan data.

This method was tested on two different brands of publically available infant formula powder. Figure 2 shows the extracted quantifier ion chromatograms for the water-soluble vitamins detected in one brand of infant formula powder.

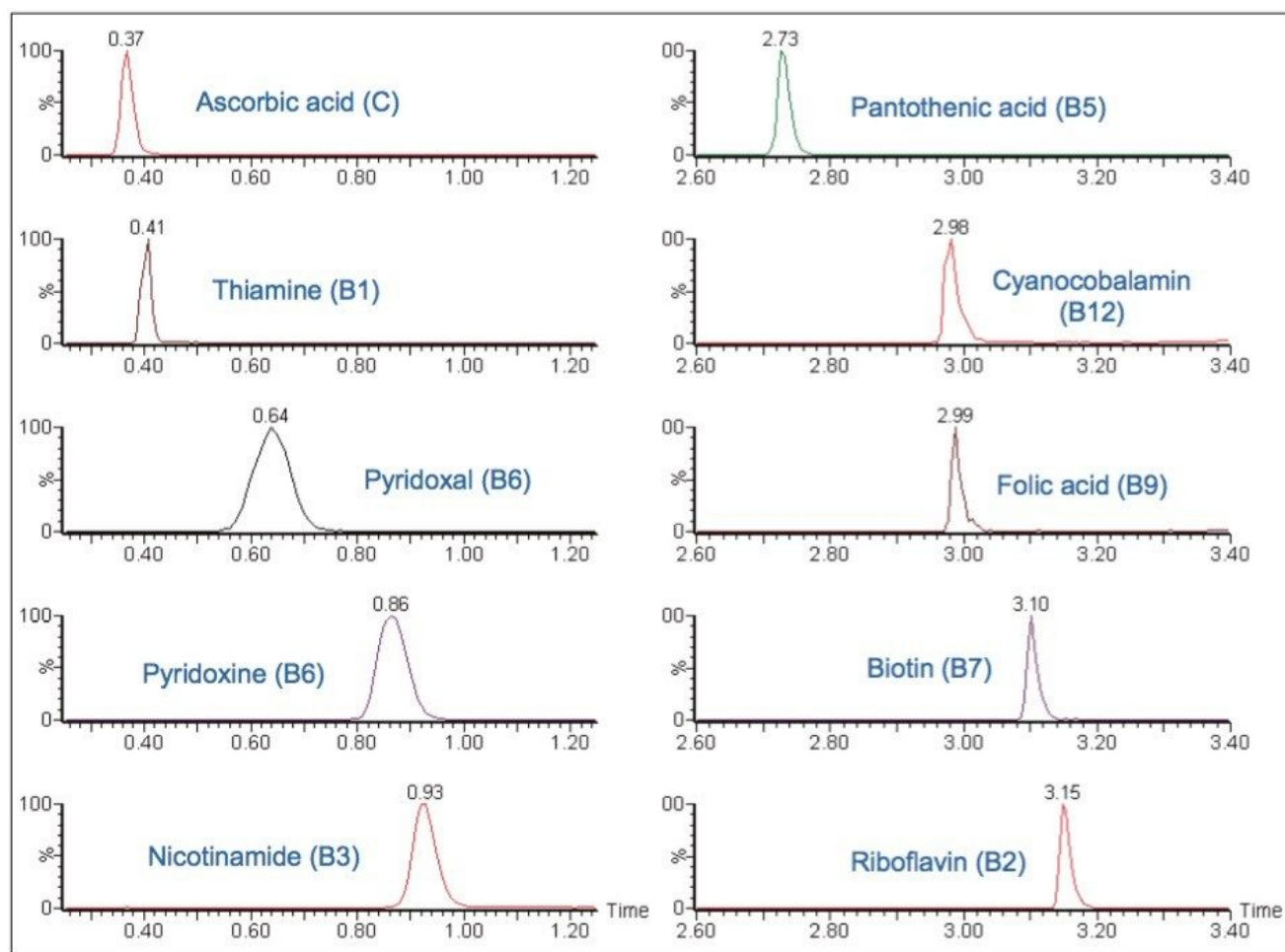


Figure 2. Extracted ion chromatograms from the analysis of infant formula powder, showing the quantifier transition.

Recovery and reproducibility

Recovery was determined by comparing the pre-extracted spiked samples with the post-extracted spiked samples. Experiments were performed for two different brands of infant formula powders on two different days, with six replicates being performed on each day.

For both brands of infant formulas, extraction recoveries of $\geq 90\%$ were generally achieved for most of the compounds, with the exception of pyridoxal, folic acid, and riboflavin-5'-phosphate, whose lower recoveries could most probably be due to their poor solubility.

Despite infant formula powder being a very complex matrix, RSDs of less than 5% were attained for six replicates. This shows high reproducibility and robustness of the proposed solution.

Analyte		Spiked level (ng/g)	Formula A		Formula B	
			Average Recovery (%)	%RSD (n=6)	Average Recovery (%)	%RSD (n=6)
Ascorbic acid	C	1000	100.4	1.1	99.4	1.1
Thiamine	B1	10	98.7	1.6	118.6	2.2
Nicotinic acid	B3	10	90.3	4.0	96.4	4.2
Pyridoxal	B6	10	83.5	1.2	88.0	1.1
Pyridoxine	B6	10	98.9	0.7	100.1	0.7
Nicotinamide	B3	10	101.6	0.5	98.7	0.6
Pantothenic acid	B5	10	104.2	1.9	112.6	0.8
Cyanocobalamin	B12	10	111.4	1.0	106.2	0.9
Folic acid	B9	100	86.6	3.7	82.0	2.4
Riboflavin-5'-phosphate	B2	100	77.6	2.2	75.1	2.8
Biotin	B7	10	95.4	2.2	98.7	2.4
Riboflavin	B2	10	102.7	1.8	103.3	2.2

Table 2. Recoveries and % RSDs of water-soluble vitamin compounds in pre-extracted spiked samples from two infant formula products.

Linearity and quantitation

Linear dynamic range, sensitivity, and suitability of the linear model were evaluated by applying the standard-addition method. An advantage of the standard-addition method is the avoidance of the evaluation of the matrix effect, responsible for signal ion suppression.

Excellent linearities were observed with correlation coefficients ≥ 0.99 , as shown in Table 3, for all of the analytes in infant formula tested, over wide concentration ranges: 10 to 10000 ng/mL for ascorbic acid (C); 0.1 to 10.0 ng/mL for cyanocobalamin (B12); and 1 to 100 ng/mL for the rest of the analytes. Calibration curves of ascorbic acid (C), and cyanocobalamin (B12), based on standard-addition in one brand of infant formula powder are shown in Figure 3.

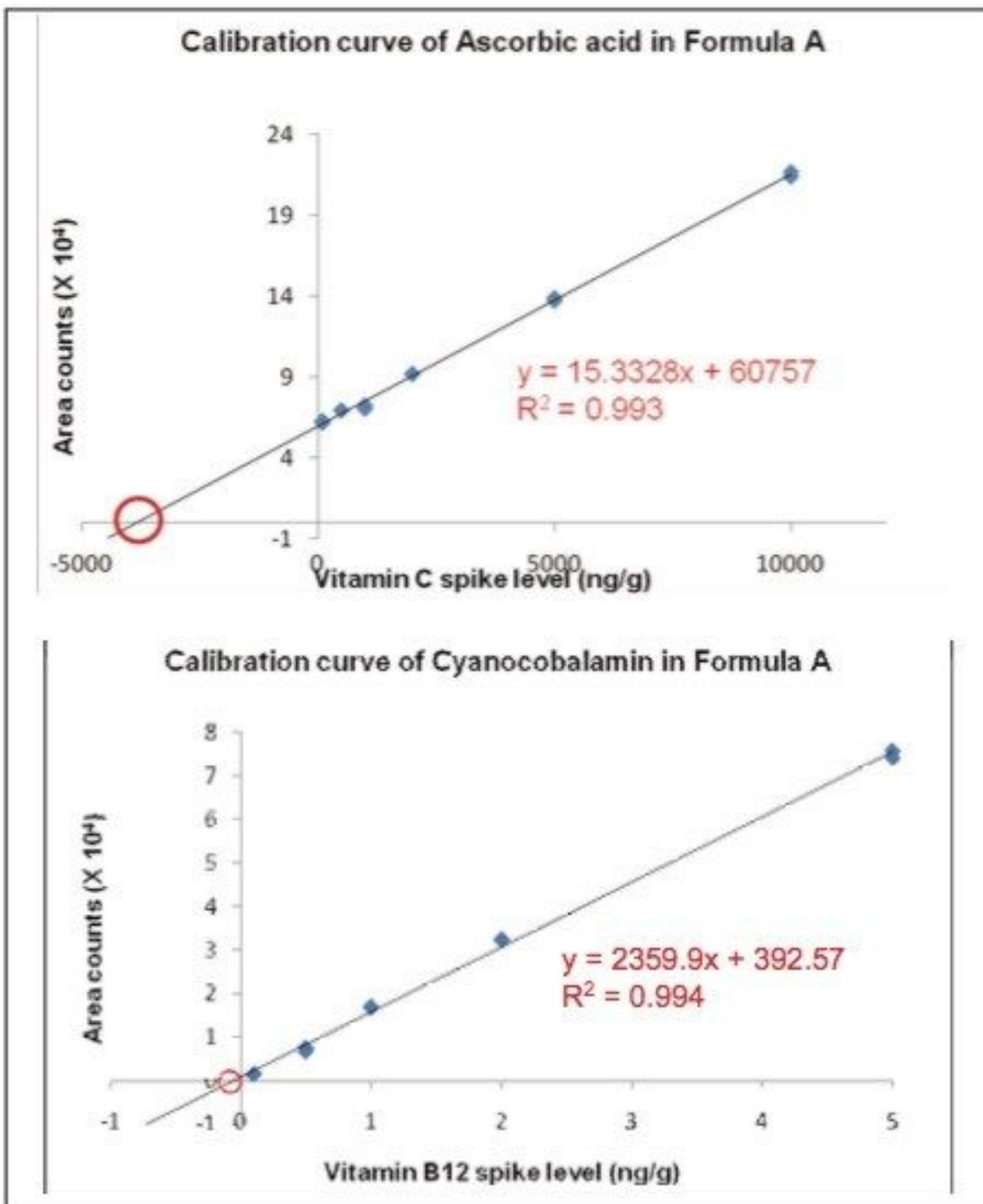


Figure 3. Calibration curves of ascorbic acid and vitamin B12 in Formula A using standard-addition.

The calibration curve was extrapolated, and the absolute value of the x-intercept showed the concentration of the analyte in the infant formula, after taking into account the dilution factor (50 x) that was used. The sensitivity of the Xevo TQ MS allowed us to dilute the sample to reduce matrix effects, while still detecting the target compounds with confidence.

Analyte		R ²	Calculated concentration (ng/g)
Ascorbic acid	C	0.993	198130
Thiamine	B1	0.990	1778
Nicotinic acid	B3	0.999	Not present
Pyridoxal	B6	0.996	205
Pyridoxine	B6	0.998	2354
Nicotinamide	B3	0.992	14201
Pantothenic acid	B5	0.994	8113
Cyanocobalamin	B12	0.994	8.3
Folic acid	B9	0.993	152
Riboflavin-5'-phosphate	B2	0.997	Not present
Biotin	B7	0.997	65
Riboflavin	B2	0.992	1143

Table 3. Correlation coefficients (R^2) for calibration curves constructed, based on the quantifier transitions, and calculated concentrations of water-soluble vitamins in Formula A.

Conclusion

A rapid 5-minute method using ACQUITY UPLC with Xevo TQ MS in positive ESI ionization mode was developed for the simultaneous analysis of 12 water-soluble vitamin compounds. This method replaces individual, lengthy methods for vitamin analysis. By combining separate vitamin analyses into a single run, laboratories can increase sample analysis throughput, reduce solvent consumption, and decrease their operational costs.

With the sensitivity of the Xevo TQ MS, it is possible to detect target compounds at low concentrations (particularly cyanocobalamin), in a very complex matrix, such as infant formula powder. With the low limit of

quantification achievable on the Xevo TQ MS, samples can be diluted to reduce matrix effects.

RADAR Technology allows for monitoring of matrix interferences, impurities, and degradants in samples, while accurately quantifying target compounds. This allows analysts to make informed decisions when assessing matrix effects, and enables a true assessment of whether matrix effects are likely to be present.

IntelliStart Technology simplifies system setup and MRM methods development, ensuring scientists of all levels can operate the instrument quickly and confidently, and start generating reproducible UPLC-MS/MS data of the highest quality.

References

1. AOAC Official Method 985.32, Microbiological Method for Analysis of Vitamin B6 (Pyridoxine, Pyridoxal, Pyridoxamine) in Ready-to-feed Milk-based Infant Formula.
2. AOAC Official Method 992.07, Microbiological Turbidimetric Method of Pantothenic Acid in Milk-based Infant Formula.

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720003694, September 2010

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