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#### 응용 자료

# Quantitative Determination of Noncovalent Protein-Ligand Interactions Automatic Chip-Based Nanoelectrospray

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# Abstract

The purpose of this experiment is to demonstrate automated nanoESI/MS analysis to determine micromolar and submicromolar dissociation constants as well as to measure the solution binding constants for the Ribonuclease A (RNase) complexes with cytidilic acid ligands.

#### Benefits

- · Allows extended acquisition time for better data quality
- · Increases sample throughput
- · Improves spray stability and reproducibility
- · Automates nanoelectrospray with one time spray optimization
- · No carryover

# Introduction

### Automated Chip-Based Nanoelectrospray

Advion BioSciences, Inc. (Ithaca, NY) has developed a method and demonstrated the capabilities of the NanoMate System for quantitative determination of noncovalent interactions between proteins and ligands. The NanoMate 100 is a chip-based automated nanoelectrospray ionization system for mass spectrometry, and is readily integrated with the Waters Micromass Q-Tof micro.

Investigations of noncovalent protein-ligand interactions by nanoelectrospray ionization mass spectrometry (nanoESI/MS) are of great interest because of their relevance to molecular recognition and to combinatory ligand library searching. This application note from Advion Biosciences introduces an experiment where automated nano ESI/MS analysis has been used to determine micromolar and submicromolar dissociation constants as well as to measure the solution biding constants for the Ribonuclease A (RNase) complexes with cytidilic acid ligands.



Waters Micromass Q-Tof micro Mass Spectrometer and the Advion NanoMate 100.

# Experimental

## Determination of Noncovalent Protein-Ligand Interactions

RNase complexed with cytidine 2'-monophosphate and cytidine triphosphate (see Figure 1), a well characterized model system, was used to demonstrate the method.

#### **Titration Experiments**

RNase protein was maintained at 10  $\mu$ M and 4  $\mu$ M, respectively, in 10 mM ammonium acetate pH 6.8 for titration of 2'-CMP (1 to 20  $\mu$ M) and CTP (1 to 8  $\mu$ M), respectively. The solutions were then incubated at room temperature for 15 minutes prior to MS analysis.

$$Kd = \frac{[R] * [L]}{[RL]} = \frac{[R] * ([Li] - [RL])}{[RL]}$$
$$[RL]/[R] = 1/Kd * ([Li] - [RL])$$

#### **Competitive Binding Experiments**

Equimolar solutions of 2'-CMP and CTP (4  $\mu$ M) were mixed with 4  $\mu$ M of RNase in 10 mM ammonium acetate pH 6.8. The solutions were then incubated at room temperature for 15 minutes prior to nanoelectrospray MS analysis.

 $Kd_{RL1} = [R] * ([R] + [RL_2]) / [RL_1]$  $Kd_{RL2} = [R] * ([R] + [RL_1]) / [RL_2]$ 

### NanoESI/MS Analytical Conditions

Sample size:	3 µL	
Flow rate:	100 nL/minute	
Spray voltage:	1.5 kV	
Pressure:	0.3 psi	
Acquisition time:	2 minutes	
Instrumentation:	NanoMate100 with ESI Chip Micromass Q-Tof micro	
Sample cone voltage:	30 V	

# **Results and Discussion**

Each ligand was detected using the NanoMate100 System (Figures 2 and 3) and as a result titration and competitive binding experiments were performed (Figure 4). The results presented are in agreement with previously published results of circular dichroism (CD).<sup>1</sup>



Figure 1. Protein and ligand structures.



Figure 2. NanoESI mass spectra.



Figure 3. Deconvoluted mass spectra of the RNase-Ligand complexes.



Figure 4. Titration assay for RNase (10mM) with 2'-CMP.

	Kd (μM)		
Ligand	Titration experiment		Competitive binding
	Ave. of individual points	Plot	experiment
Cytidine 2'- monophosphate (2'-CMP)	1.71 ± 0.33	2.00 ± 0.43	$2.3 \pm 0.4$
Cytidine triphosphate (CTP)	$0.80 \pm 0.2$	0.74 ± 0.3	0.75 ± 0.4

Table 1. Summary of binding assay for RNase and cytidine nucleotideligands using automated NanoESI/MS.

## Conclusion

The NanoMate100 System can be used to determine micromolar and submicromolar dissociation constants. In addition, an automated nanoESI/MS method can be used to measure solution binding constants for the RNase complexes with cytidilic acid ligands.

# References

- 1. Jones, C.L.; Fish, F.; Muccio, D.D. Anal BioChem 2002, 302, 184–190.
- 2. Application note based on Zhang, S.; Van Pelt, C.K.; Wilson, D.B. Anal Chem 2003, 75, 3010-3018.

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